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(54) Title: CUTINASE VARIANTS

(57) Abstract

Variants of fungal cutinases have improved thermostability. The variants comprise substitution of one or more amino acid residues near the N-terminal in the amino acid sequence or in the three-dimensional structure of the cutinase.

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CUTINASE VARIANTS

FIELD OF THE INVENTION

The present invention relates to a cutinase variant, more particularly to a cutinase variant having improved thermostability. The invention also relates to a DNA sequence encoding the variant, a vector comprising the DNA sequence, a transformed host cell harboring the DNA sequence or the vector, to a method of producing the variant, and to use of the variant.

BACKGROUND OF THE INVENTION

Cutinases are lipolytic enzymes capable of hydrolyzing the substrate cutin.

10 Cutinases are known from various fungi (P.E. Kolattukudy in "Lipases", Ed. B. Borgström and H.L. Brockman, Elsevier 1984, 471-504). The amino acid sequence and the crystal structure of a cutinase of *Fusarium solani pisi* have been described (S. Longhi et al., Journal of Molecular Biology, 268 (4), 779-799 (1997)). The amino acid sequence of a cutinase from *Humicola insolens* has also been published (US 5,827,719).

A number of variants of the cutinase of *Fusarium solani pisi* have been published: WO 94/14963; WO 94/14964; Appl. Environm. Microbiol. 64, 2794-2799, 1998; Proteins: Structure, Function and Genetics 26, 442-458, 1996; J. of Computational Chemistry 17, 1783-1803, 1996; Protein Engineering 6, 157-165, 1993; Proteins: Structure, Function, and Genetics 33, 253-264, 1998; J. of Biotechnology 66, 11-26, 1998; Biochemistry 35, 398-410, 1996.

Fungal cutinases may be used in the enzymatic hydrolysis of cyclic oligomers of poly(ethylene terephthalate), e.g. in the finishing of yarn or fabric from poly(ethylene terephthalate) fibers (WO 97/27237). However, it is desirable to improve the thermostability of known fungal cutinases to allow a higher process temperature.

SUMMARY OF THE INVENTION

The inventors have found certain variants of fungal cutinases having improved thermostability.

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Accordingly, the invention provides a variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which is located:

- a) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
- b) within 20 positions from the N-terminal amino acid.

The invention also provides a DNA sequence encoding the variant, an expression vector comprising the DNA sequence, a transformed host cell harboring the DNA sequence or the expression vector, a method of producing the variant, processes using the variant and a detergent composition comprising the variant.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 gives the coordinates for the 3D structure of the cutinase of *H. insolens*.

Fig. 2 is a computer model showing the three-dimensional structures of the cutinases from *F. solani pisi* (left) and *H. insolens* (right). Different colors have been used to identify the N-terminal amino acid and zones of 12 Å and 17 Å diameter around this.

Figs. 3-6 illustrate the hydrolysis of c3ET. Details are given in the Examples.

DETAILED DESCRIPTION OF THE INVENTION

20 Fungal cutinase

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The parent cutinase is a fungal cutinase, such as a filamentous fungal cutinase, e.g. native to a strain of *Humicola* or *Fusarium*, specifically *H. insolens* or *F. solani pisi*, more specifically *H. insolens* strain DSM 1800.

The amino acid sequence of the cutinase of *H. insolens* strain DSM 1800 and the DNA sequence encoding it are shown as SEQ ID NO: 2 and SEQ ID NO: 1 of US 5,827,719. The numbering system used herein for the *H. insolens* cutinase is based on the mature peptide, as shown in said SEQ ID NO: 2.

The amino acid sequence of the cutinase of *F. solani pisi* is shown as the mature peptide in Fig. 1D of WO 94/14964. The numbering system used herein for

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the F. solani pisi cutinase is that used in WO 94/14964; it includes the prosequence shown in said Fig. 1D; thus, the mature cutinase is at positions 16-214.

The parent cutinase may have an amino acid sequence which is at least 50 % (particularly at least 70 % or at least 80 %) homologous to the cutinase of H. inso-5 lens strain DSM 1800. The parent cutinase may particularly be one that can be aligned with the cutinase of H. insolens strain DSM 1800.

Nomenclature for amino acids and alterations

The specification and claims refer to amino acids by their one-letter codes. A particular amino acid in a sequence is identified by its one-letter code and its posi-10 tion, e.g. Q1 indicates Gln (glutamine at position 1, i.e. at the N-terminal.

The nomenclature used herein for defining substitutions is basically as described in WO 92/05249. Thus, R51P indicates substitution of R (Arg) at position 51 with P (Pro).

Homology and alignment

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For purposes of the present invention, the degree of homology may be suitably determined according to the method described in Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-45, with the following settings for polypeptide sequence comparison: GAP creation penalty of 3.0 and GAP extension penalty of 0.1. The determination may be done by means of a computer program 20 known such as GAP provided in the GCG program package (Program Manual for the Wisconsin Package, Version 8, August 1994, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711).

Two given sequences can be aligned according to the method described in Needleman (supra) using the same parameters. This may be done by means of the 25 GAP program (supra).

Three-dimensional structure of cutinase

The structure of the cutinase of H. insolens was solved in accordance with the principle for X-ray crystallographic methods as given, for example, in X-Ray Structure Determination, Stout, G.K. and Jensen, L.H., John Wiley & Sons, Inc. NY, 30 1989. The structural coordinates for the solved crystal structure at 2.2 Å resolution

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using the isomorphous replacement method are given in Fig. 1 in standard PDB format (Protein Data Bank, Brookhaven National Laboratory, Brookhaven, CT).

The structure of the cutinase of *F. solani pisi* is described in Martinez et al. (1992) Nature 356, 615-618. The 3D structures of the cutinases of *F. solani pisi* and *H. insolens* are compared as a computer model in Fig. 2.

It should be noted that the overall three-dimensional structures of fungal cutinases are very similar and have been shown by X-ray crystallography to be highly homologous. The similarities between the cutinases from *F. solani pisi* and *H. insolens* are clearly apparent from the computer model in Fig. 2. Therefore, modifications of the type indicated for one fungal cutinase will also be functional for other fungal cutinases.

Substitution near N-terminal

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The variant of the invention has one or more amino acid substitutions in the vicinity of the N-terminal. The substitution is within a distance of 17 Å (e.g. within 12 Å) and/or within 20 positions (e.g. within 15 positions) of the N-terminal. The distance from the N-terminal is to be calculated between the Cα atom of the amino acids, and is calculated from an amino acid in a crystal structure (i.e. visible in the X-ray structure).

In the cutinase of *H. insolens* strain DSM 1800, the two N-terminal amino acids (Q1 and L2. i.e. Gln and Leu at positions 1 and 2) are not visible in the X-ray structure, so the distance is to be calculated from amino acid G3. Amino acids within 17 Å include positions 3-12, 18, 20-60, 62-64, 82, 85-86, 100-108, 110-111, 130-132, 174, 176-182, 184-185, 188, and 192. Those within 12 Å include positions 3-8, 25 22-27, 30-47, 53-59, 102, 177, and 180-181.

In the cutinase of *F. solani pisi*, the N-terminal amino acid G17 is visible in the X-ray structure. Amino acids within 17 Å include positions 17-26, 34-75, 77-79, 101, 115, 117-119, 147, 191-197, 199-200, and 203. Those within 12 Å include positions 17-22, 38, 40, 45-58, 60, 65, and 70-72.

The variants of the invention have improved thermostability compared to the parent enzyme. The thermostability may be determined from the denaturation tem-

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perature by DSC (differential scanning calorimetry), e.g. as described in an example, e.g. at pH 8.5 with a scan rate of 90 K/hr. The variants may have a denaturation temperature which is at least 5°C higher than the parent enzyme.

The total number of substitutions in the above regions is typically 1-10, e.g. 1-5 substitutions in the above regions. In addition, the cutinase variant of the invention may optionally include other modifications of the parent enzyme, typically 10 or fewer, e.g. 5 or fewer alterations (substitutions, deletions or insertions) outside of the above regions. Thus, the total amino acid sequence of the variant typically 1-20, e.g. 1-10 alterations compared to the parent cutinase.

10 Solvent accessible surface

One or more of the substitutions may be made at an exposed amino acid residue, i.e. an amino acid residue having a solvent accessible surface. This can be calculated by the "dssp" program (version October 1988) described in W. Kabsch and C. Sander, Biopolymers, 22 (1983) pp. 2577-2637.

In the cutinase of *H. insolens* strain DSM 1800, the following amino acids lie within 17 Å of G3 at the N-terminal and have a solvent accessible surface greater than 0: 3-12, 18, 26-33, 36-38, 40-45, 47-56, 59-60, 62-64, 82, 85-86, 104-105, 174, 176-179, 181-182, 192.

Specific substitutions

The substitution near the N-terminal may specifically be one that increases the electrical charge, i.e. a substitution of a negatively charged amino acid with a neutral or positively charged amino acid or substitution of a neutral amino acid with a positively charged amino acid. Thus, a negative amino acid residue at a position corresponding to position E6, E10, E30, E47 D63, E82 and/or E179 in the cutinase of Humicola insolens strain DSM 1800 may be substituted by a neutral or positive amino acid, e.g. R, K, Y, H, Q or N. Some specific substitutions are those corresponding to E6Q/N, E10Q/N, E47K/R or E179Q/N. Also, a neutral amino acid residue at a position corresponding to N7, S11, N44 or N52 in the H. insolens cutinase may be substituted by a positive amino acid (R, K or H).

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Another example of a substitution near the N-terminal is substitution with a Pro residue, e.g. a substitution corresponding to A14P or R51P in the cutinase of *Humicola insolens* strain DSM 1800.

Specific variants

The following are some examples of variants in the *H. insolens* cutinase. Corresponding variants may be made on the basis of other parent cutinases.

R51P

E6N/Q+ L1381

A14P+ E47K

10 E47K

E179N/Q

E6N/Q+ E47K+ R51P

A14P+ E47K+ E179N/Q

E47K+ E179N/Q

15 E47K+ D63N

E6N/Q+ E10N/Q+ A14P+ E47K+ R51P+ E179N/Q

E6N/Q+ A14P+ E47K+ R51P+ E179N/Q

Q1P+ L2V+ S11C+ N15T+ F24Y+ L46I+ E47K

Use of cutinase variant

The cutinase variant of the invention may be used, e.g., for the enzymatic hydrolysis of cyclic oligomers of poly(ethylene terephthalate), such as cyclic tri(ethylene terephthalate), abbreviated as c3ET.

In particular, this may be used to remove such cyclic oligomers from polyester containing fabric or yarn by treating the fabric or yarn with the cutinase variant, optionally followed by rinsing the fabric or yarn with an aqueous solution having a pH in the range of from about pH 7 to about pH 11. The treatment of polyester is conveniently carried out above the glass transition temperature of c3ET (about 55°C) and below the glass transition temperature of polyester (about 70°C). Thus, the treatment may suitably be carried out at 50-80°C, e.g. at 60-75°C. The process may be carried out in analogy with WO 97/27237.

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The cutinase variant may be used to treat polyester-containing textile. e.g. PET (polymer of ethyleneglycol and terephthalic acid), P3GT (polymer of 1,3-propanediol and terephthalic acid) or a polyester/cotton blend. The treatment may provide benefits to the polyester textile such as improved wear and comfort, increased water permeability, reduced antistatic behavior, improve handle and softness, changed redeposition characteristics and/or color clarification.

The cutinase variant may be used to improve the functional finish of a PET-containing yarn or fabric by a treatment with the cutinase variant, followed by a treatment with a finishing agent such as a softener, an anti-crease resin, an anti-static agent, an anti-soiling agent or agents to impair wrinkle-free, permanent press ior fire resistance effects. The treatment with the cutinase variant may increase the number of functional groups in the surface, and this can be used to attach the functional finish. Examples of finishing agents are described in "SENSHOKU SIAGEKAKO BENRAN" published 1998-10-15 by Nihon Seni Sentaa KK.

The cutinase variant of the invention is also useful in detergents, where it may be incorporated to improve the removal of fatty soiling, as described in WO 94/03578 and WO 94/14964. The addition of the cutinase variant to laundry detergent may reduce malodor from cloth which is accumulated during several wash/wear-cycles.

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The cutinase variant may also be used for degradation and recycling of polyes-20 ter such as polycaprolactone (PCL), poly-ethyleneglycol-terephthalate (PET), polylactic acid, polybutylenesuccinate, and poly(hydroxybutiric acid)-co-(hydroxyvaleric acid), e.g. film and bottles, e.g. as described in JP-A 5-344897.

The cutinase variant may also be used for other known applications of lipases and cutinases, for example, in the baking industry (e.g. as described in WO 94/04035 and EP 585988), in the papermaking industry (e.g. for pitch removal, see EP 374700), and in the leather, wool and related industries (e.g. for degreasing of animal hides, sheepskin or wool), and for other applications involving degreasing/defatting. It may be used in immobilized form in the fat and oil industry, as a catalyst in organic synthesis (e.g. esterification, transesterification or ester hydrolysis reactions).

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Dyeing polyester

The invention provides a process for dyeing polyester fabric or yarn. In this process, the fabric or yarn is first treated with a cutinase, e.g. 12-48 hours at 50-70°C or 65-70°C, pH 7-10, followed by dyeing with dye, e.g. a reactive dye, a disperse dye or a cationic dye. The reactive dye may be one that reacts with OH or COOH groups, e.g. having the structure Chromophore-NHPh-SO₂CH₂CH₂OSO₃Na. The dyeing may be conducted at 40-80°C, e.g. for 20-60 minutes.

The cutinase may be a thermostable cutinase having a thermal denaturation temperature, T_d, at pH 8.5 which is at least 5° higher than the parent cutinase, e.g. 7-10° higher, e.g. a value of 65°C or higher. The measurement may be made by DSC as described in an Example of this specification.

Surfactant

In the treatment of fabric or yarn, a conventional wetting agent and/or a dispersing agent may be used to improve the contact with the enzyme. The wetting agent may be a nonionic surfactant, e.g. an ethoxylated fatty alcohol. A very useful wetting agent is an ethoxylated and propoxylated fatty acid ester such as Berol 087 (product of Akzo Nobel, Sweden).

The dispersing agent may suitably be selected from nonionic, anionic, cationic, ampholytic or zwitterionic surfactants. More specifically, the dispersing agent may be selected from carboxymethylcellulose, hydroxypropylcellulose, alkyl aryl sulfonates, long-chain alcohol sulfates (primary and secondary alkyl sulfates), sulfonated olefins, sulfated monoglycerides, sulfated ethers, sulfosuccinates, sulfonated methyl ethers, alkane sulfonates, phosphate esters, alkyl isothionates, acylsarcosides, alkyltaurides, fluorosurfactants, fatty alcohol and alkylphenol condensates, fatty acid condensates, condensates of ethylene oxide with an amine, condensates of ethylene oxide with an amide, sucrose esters, sorbitan esters, alkyloamides, fatty amine oxides, ethoxylated monoamines, ethoxylated diamines, alcohol ethoxylate and mixtures thereof. A very useful dispersing agent is an alcohol ethoxylate such as Berol 08 (product of Akzo Nobel, Sweden).

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Methods for preparing cutinase variants

The cutinase variant of the invention can be prepared by methods known in the art, e.g. as described in WO 94/14963 or WO 94/14964 (Unilever). The following describes methods for the cloning of cutinase-encoding DNA sequences, followed by methods for generating mutations at specific sites within the cutinase-encoding sequence.

Cloning a DNA sequence encoding a cutinase

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The DNA sequence encoding a parent cutinase may be isolated from any cell or microorganism producing the cutinase in question, using various methods well known in the art. First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the cutinase to be studied. Then, if the amino acid sequence of the cutinase is known, labeled oligonucleotide probes may be synthesized and used to identify cutinase-encoding clones from a genomic library prepared from the organism in question. Alternatively, a labeled oligonucleotide probe containing sequences homologous to another known cutinase gene could be used as a probe to identify cutinase-encoding clones, using hybridization and washing conditions of lower stringency.

Yet another method for identifying cutinase-encoding clones would involve inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming cutinase-negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto agar containing a substrate for cutinase (i.e. maltose), thereby allowing clones expressing the cutinase to be identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g. the phosphoroamidite method described S.L. Beaucage and M.H. Caruthers, (1981), Tetrahedron Letters 22, p. 1859-1869, or the method described by Matthes et al., (1984), EMBO J. 3, p. 801-805. In the phosphoroamidite method, oligonucleotides are synthesized, e.g. in an automatic DNA synthesizer, purified, annealed, ligated and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed genomic and synthetic origin, mixed synthetic and cDNA origin or mixed genomic and cDNA origin, prepared by

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ligating fragments of synthetic, genomic or cDNA origin (as appropriate, the fragments corresponding to various parts of the entire DNA sequence), accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in 5 US 4,683,202 or R.K. Saiki et al., (1988), Science 239, 1988, pp. 487-491.

Site-directed mutagenesis

Once a cutinase-encoding DNA sequence has been isolated, and desirable sites for mutation identified, mutations may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired 10 mutation sites. In a specific method, a single-stranded gap of DNA, the cutinaseencoding sequence, is created in a vector carrying the cutinase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the construct is ligated using T4 ligase. A specific example 15 of this method is described in Morinaga et al., (1984), Biotechnology 2, p. 646-639. US 4,760,025 discloses the introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette. However, an even greater variety of mutations can be introduced at any one time by the Morinaga method, because a multitude of oligonucleotides, of various lengths, can be introduced.

Another method for introducing mutations into cutinase-encoding DNA sequences is described in Nelson and Long, (1989), Analytical Biochemistry 180, p. 147-151. It involves the 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the PCR reactions. From the PCR-generated fragment, a DNA fragment 25 carrying the mutation may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

Expression of cutinase variants

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According to the invention, a DNA sequence encoding the variant produced by methods described above, or by any alternative methods known in the art, can be 30 expressed, in enzyme form, using an expression vector which typically includes con-

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trol sequences encoding a promoter, operator, ribosome binding site. translation initiation signal, and, optionally, a repressor gene or various activator genes.

Expression vector

The recombinant expression vector carrying the DNA sequence encoding a cutinase variant of the invention may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. The vector may be one which, when introduced into a host cell, is integrated into the host cell genome and replicated to-10 gether with the chromosome(s) into which it has been integrated. Examples of suitable expression vectors include pMT838.

Promoter

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In the vector, the DNA sequence should be operably connected to a suitable promoter sequence. The promoter may be any DNA sequence which shows tran-15 scriptional activity in the host cell of choice and may be derived from genes encoding proteins either homologous or heterologous to the host cell.

Examples of suitable promoters for directing the transcription of the DNA sequence encoding a cutinase variant of the invention, especially in a bacterial host. are the promoter of the lac operon of E.coli, the Streptomyces coelicolor agarase 20 gene dagA promoters, the promoters of the Bacillus licheniformis α-amylase gene (amyL), the promoters of the Bacillus stearothermophilus maltogenic amylase gene (amyM), the promoters of the Bacillus amyloliquefaciens α -amylase (amyQ), the promoters of the Bacillus subtilis xylA and xylB genes etc. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding A. 25 oryzae TAKA amylase, the TPI (triose phosphate isomerase) promoter from S. cerevisiae (Alber et al. (1982), J. Mol. Appl. Genet 1, p. 419-434, Rhizomucor miehei aspartic proteinase, A. niger neutral α -amylase, A. niger acid stable α -amylase, A. niger glucoamylase, Rhizomucor miehei lipase, A. oryzae alkaline protease, A. oryzae triose phosphate isomerase or A. nidulans acetamidase.

Expression vector

The expression vector of the invention may also comprise a suitable transcription terminator and, in eukaryotes, polyadenylation sequences operably connected to the DNA sequence encoding the α -amylase variant of the invention. Ter-5 mination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and pIJ702.

The vector may also comprise a selectable marker, e.g. a gene the product of which complements a defect in the host cell, such as the dal genes from B. subtilis or B. licheniformis, or one which confers antibiotic resistance such as ampicillin, kanamycin, chloramphenicol or tetracyclin resistance. Furthermore, the vector may comprise Aspergillus selection markers such as amdS, argB, niaD and sC, a marker 15 giving rise to hygromycin resistance, or the selection may be accomplished by cotransformation, e.g. as described in WO 91/17243.

The procedures used to ligate the DNA construct of the invention encoding a cutinase variant, the promoter, terminator and other elements, respectively, and to insert them into suitable vectors containing the information necessary for replication, 20 are well known to persons skilled in the art (cf., for instance, Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, 1989).

Host Cells

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The cell of the invention, either comprising a DNA construct or an expression vector of the invention as defined above, is advantageously used as a host cell in 25 the recombinant production of a cutinase variant of the invention. The cell may be transformed with the DNA construct of the invention encoding the variant, conveniently by integrating the DNA construct (in one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA con-30 structs into the host chromosome may be performed according to conventional

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methods, e.g. by homologous or heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

The cell of the invention may be a cell of a higher organism such as a mam-5 mal or an insect, but is preferably a microbial cell, e.g. a bacterial or a fungal (including yeast) cell.

Examples of suitable bacteria are Gram positive bacteria such as *Bacillus subtilis*, *Bacillus licheniformis*, *Bacillus lentus*, *Bacillus brevis*, *Bacillus stearothermophilus*, *Bacillus alkalophilus*, *Bacillus amyloliquefaciens*, *Bacillus coagulans*, *Bacillus coagulans*, *Bacillus circulans*, *Bacillus lautus*, *Bacillus megaterium*, *Bacillus thuringiensis*, or *Streptomyces lividans* or *Streptomyces murinus*, or gramnegative bacteria such as *E.coli*. The transformation of the bacteria may, for instance, be effected by protoplast transformation or by using competent cells in a manner known *per se*.

The yeast organism may favorably be selected from a species of *Saccharo-*15 myces or *Schizosaccharomyces*, e.g. *Saccharomyces* cerevisiae.

The host cell may also be a filamentous fungus e.g. a strain belonging to a species of Aspergillus, most preferably Aspergillus oryzae or Aspergillus niger, or a strain of Fusarium, such as a strain of Fusarium oxysporium, Fusarium graminearum (in the perfect state named Gribberella zeae, previously Sphaeria zeae, synonym with Gibberella roseum and Gibberella roseum f. sp. cerealis), or Fusarium sulphureum (in the prefect state named Gibberella puricaris, synonym with Fusarium trichothecioides, Fusarium bactridioides, Fusarium sambucium, Fusarium roseum, and Fusarium roseum var. graminearum), Fusarium cerealis (synonym with Fusarium crokkwellnse), or Fusarium venenatum.

In a preferred embodiment of the invention the host cell is a protease deficient or protease minus strain.

This may for instance be the protease deficient strain *Aspergillus oryzae* JaL 125 having the alkaline protease gene named "alp" deleted. This strain is described in WO 97/35956 (Novo Nordisk).

Filamentous fungi cells may be transformed by a process involving protoplast formation and transformation of the protoplasts followed by regeneration of the cell wall in a manner known per se. The use of *Aspergillus* as a host micro-organism

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is described in EP 238 023 (Novo Nordisk A/S), the contents of which are hereby incorporated by reference.

Production of cutinase variant by cultivation of transformant

The invention relates, *inter alia*, to a method of producing a cutinase variant of the invention, which method comprises cultivating a host cell under conditions conducive to the production of the variant and recovering the variant from the cells and/or culture medium.

The medium used to cultivate the cells may be any conventional medium suitable for growing the host cell in question and obtaining expression of the cutinase variant of the invention. Suitable media are available from commercial suppliers or may be prepared according to published recipes (e.g. as described in catalogues of the American Type Culture Collection).

The cutinase variant secreted from the host cells may conveniently be recovered from the culture medium by well-known procedures, including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulphate, followed by the use of chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

Expression of variant in plants

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The present invention also relates to a transgenic plant, plant part or plant cell which has been transformed with a DNA sequence encoding the variant of the invention so as to express and produce this enzyme in recoverable quantities. The enzyme may be recovered from the plant or plant part. Alternatively, the plant or plant part containing the recombinant enzyme may be used as such.

The transgenic plant can be dicotyledonous or monocotyledonous, for short a dicot or a monocot. Examples of monocot plants are grasses, such as meadow grass (blue grass, Poa), forage grass such as festuca, lolium, temperate grass, such as Agrostis, and cereals, e.g. wheat, oats, rye, barley, rice, sorghum and maize (corn).

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Examples of dicot plants are tobacco, legumes, such as lupins, potato, sugar beet, pea, bean and soybean, and cruciferous (family Brassicaceae), such as cauliflower, oil seed rape and the closely related model organism Arabidopsis thaliana.

Examples of plant parts are stem, callus, leaves, root, fruits, seeds, and tubers. In the present context, also specific plant tissues, such as chloroplast, apoplast, mitochondria, vacuole, peroxisomes and cytoplasm are considered to be a plant part. Furthermore, any plant cell, whatever the tissue origin, is considered to be a plant part.

Also included within the scope of the invention are the progeny of such plants, plant parts and plant cells.

The transgenic plant or plant cell expressing the variant of the invention may be constructed in accordance with methods known in the art. In short the plant or plant cell is constructed by incorporating one or more expression constructs encoding the enzyme of the invention into the plant host genome and propagating the resulting modified plant or plant cell into a transgenic plant or plant cell.

Conveniently, the expression construct is a DNA construct which comprises a gene encoding the enzyme of the invention in operable association with appropriate regulatory sequences required for expression of the gene in the plant or plant part of choice. Furthermore, the expression construct may comprise a selectable marker useful for identifying host cells into which the expression construct has been integrated and DNA sequences necessary for introduction of the construct into the plant in question (the latter depends on the DNA introduction method to be used).

The choice of regulatory sequences, such as promoter and terminator sequences and optionally signal or transit sequences is determined, eg on the basis of when, where and how the enzyme is desired to be expressed. For instance, the expression of the gene encoding the enzyme of the invention may be constitutive or inducible, or may be developmental, stage or tissue specific, and the gene product may be targeted to a specific tissue or plant part such as seeds or leaves. Regulatory sequences are eg described by Tague et al, Plant, Phys., 86, 506, 1988.

For constitutive expression the 35S-CaMV promoter may be used (Franck et al., 1980. Cell 21: 285-294). Organ-specific promoters may eg be a promoter from

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storage sink tissues such as seeds, potato tubers, and fruits (Edwards & Coruzzi, 1990. Annu. Rev. Genet. 24: 275-303), or from metabolic sink tissues such as meristems (Ito et al., 1994. Plant Mol. Biol. 24: 863-878), a seed specific promoter such as the glutelin, prolamin, globulin or albumin promoter from rice (Wu et al., 5 Plant and Cell Physiology Vol. 39, No. 8 pp. 885-889 (1998)), a Vicia faba promoter from the legumin B4 and the unknown seed protein gene from Vicia faba described by Conrad U. et al, Journal of Plant Physiology Vol. 152, No. 6 pp. 708-711 (1998), a promotter from a seed oil body protein (Chen et al., Plant and cell physiology vol. 39, No. 9 pp. 935-941 (1998), the storage protein napA promoter from Brassica napus, 10 or any other seed specific promoter known in the art, eg as described in WO 91/14772. Furthermore, the promoter may be a leaf specific promoter such as the rbcs promoter from rice or tomato (Kyozuka et al., Plant Physiology Vol. 102, No. 3 pp. 991-1000 (1993), the chlorella virus adenine methyltransferase gene promoter (Mitra, A. and Higgins, DW, Plant Molecular Biology Vol. 26, No. 1 pp. 85-93 (1994), 15 or the aldP gene promoter from rice (Kagaya et al., Molecular and General Genetics Vol. 248, No. 6 pp. 668-674 (1995), or a wound inducible promoter such as the potato pin2 promoter (Xu et al, Plant Molecular Biology Vol. 22, No. 4 pp. 573-588 (1993).

A promoter enhancer element may be used to achieve higher expression of the enzyme in the plant. For instance, the promoter enhancer element may be an intron which is placed between the promoter and the nucleotide sequence encoding the enzyme. For instance, Xu et al. op cit disclose the use of the first intron of the rice actin 1 gene to enhance expression.

The selectable marker gene and any other parts of the expression construct may be chosen from those available in the art.

The DNA construct is incorporated into the plant genome according to conventional techniques known in the art, including *Agrobacterium*-mediated transformation, virus-mediated transformation, micro injection, particle bombardment, biolistic transformation, and electroporation (Gasser et al, Science, 244, 1293; Potrykus, Bio/Techn. 8, 535, 1990; Shimamoto et al, Nature, 338, 274, 1989).

Presently, Agrobacterium tumefaciens mediated gene transfer is the method of choice for generating transgenic dicots (for review Hooykas & Schilperoort, 1992.

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Plant Mol. Biol. 19: 15-38), however it can also be used for transforming monocots, although other transformation methods are generally preferred for these plants. Presently, the method of choice for generating transgenic monocots is particle bombardment (microscopic gold or tungsten particles coated with the transforming DNA) of embryonic calli or developing embryos (Christou, 1992. Plant J. 2: 275-281; Shimamoto, 1994. Curr. Opin. Biotechnol. 5: 158-162; Vasil et al., 1992. Bio/Technology 10: 667-674). An alternative method for transformation of monocots is based on protoplast transformation as described by Omirulleh S, et al., Plant Molecular biology Vol. 21, No. 3 pp. 415-428 (1993).

Following transformation, the transformants having incorporated the expression construct are selected and regenerated into whole plants according to methods well-known in the art.

MATERIALS AND METHODS

Plasmids

15 pJSO026

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This is a *S. cerevisiae* expression plasmid described in WO 97/07205 and in J.S.Okkels, (1996) "A URA3-promoter deletion in a pYES vector increases the expression level of a fungal lipase in Saccharomyces cerevisiae. Recombinant DNA Biotechnology III: The Integration of Biological and Engineering Sciences, vol. 782 of the Annals of the New York Academy of Sciences).

pFuku83

This is a yeast and E. coli shuttle vector for expression of the H. insolens cutinase under the control of a TPI promoter, constructed from pJSO026.

Substrate

25 BETEB

Terephthalic acid bis(2-hydroxyethyl)ester dibenzoate is herein abbreviated as BETEB (benzoyl-ethylene-terephthalic-ethelene-benzoate). It was prepared from terephthalic acid bis (2-hydroxyethyl) ester and benzoic acid.

Lipase activity (LU)

A substrate for lipase is prepared by emulsifying tributyrin (glycerin tributyrate) using gum Arabic as emulsifier. The hydrolysis of tributyrin at 30 °C at pH 7 is followed in a pH-stat titration experiment. One unit of lipase activity (1 LU) equals the amount of enzyme capable of releasing 1 µmol butyric acid/min at the standard conditions.

Differential scanning calorimetry (DSC)

Sample and reference solutions are carefully degassed immediately prior to loading of samples into the calorimeter (reference: buffer without enzyme). Sample and reference solutions (approx. 0.5 ml) are thermally pre-equillibrated for 20 minutes at 5°C. The DSC scan is performed from 5 C to 95 C at a scan rate of approx. 90 K/hr. Denaturation temperatures are determined at an accuracy of approx. +/- 1 C. A VP-DSC from MicroCal Inc. is suitable for the experiments.

Methods

15 PCR conditions

step 1: 94° C, 120 sec.

step 2: 94° C, 60 sec

step 3: 50° C, 60 sec

step 4: 72° C, 150 sec.

Go to step 2, 35 cycles

step 5: 72° C, 480 sec.

Step 6: 4° C, for ever

EXAMPLES

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Example 1: Preparation of cutinase variants

A DNA sequence encoding *H. insolens* cutinase was obtained as described in US 5,827,719 (Novo Nordisk) and was found to have the DNA sequence shown in SEQ ID NO: 1 therein.

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Variants were prepared by localized random mutagenesis and selection of positive clones by incubation at 60°C for 1 day on BETEB plates. The BETEB plates contained 200 ml/l of 500 mM glycine buffer (pH 8.5), 1.25 g/l of BETEB (dissolved in hot ethanol) and 20 g/l of agar.

Three positive variants were isolated, and their amino acid sequence was determined. They were found to have the following modifications, compared to the parent *H. insolens* cutinase:

A14P + E47K

E47K

10 E179Q

Example 2: Site directed mutation

A variant of the *H.* insolens cutinase having the substitutions E6Q+ E47K+ R51P was prepared as follows:

A pair of PCR primers were designed so as to introduce amino acid substitu-15 tions, making use of the existed restriction enzyme sites nearby, as follows (an asterisk indicates an introduced mutation):

Upper primer: E6Q F

cgg cag ctg gga gcc atc c*ag aac

Pvu II

20 Lower primer: E47K,R51P

cgc cct gga tcc aga tgt tcg* gga tgt ggg act t*aa ggc

BamH I

PCR was run using these primers and pFukuNL83 as a template under the PCR condition described above.

The obtained PCR fragment was purified by Clontech Spincolumn and digested with *Pvu* II and *BamH* I.

The resultant fragment was gel-purified and ligated to pFukuNL83 which had been digested with the same restriction enzyme sites.

Example 3: Thermostability of cutinase variants

Variants

The thermostability was tested as described below for the *H. insolens* cutinase and the following variants thereof:

5 A14P+ E47K

E47K

E179Q

E6Q+ E47K+ R51P

A14P+ E47K+ E179Q

10 E6Q+ A14P+ E47K+ R51P+ E179Q

E6Q+ E10Q+ A14P+ E47K+ R51P+ E179Q

<u>Differential Scanning Calorimetry (DSC)</u>

Thermostability of cutinase variants was investigated by means of DSC at pH 4.5 (50 mM acetate buffer) and pH 8.5 (50mM glycyl-glycine buffer). The thermal denaturation temperature, T_d, was taken as the top of denaturation peak (major endothermic peak) in thermograms (Cp vs. T) obtained after heating of enzyme solutions at a constant programmed heating rate.

The parent cutinase was found to have T_d of 63°C at pH 8.5. Six of the above variants were found to have T_d of 70-73°C, i.e. an improvement of 7-10°.

The parent cutinase was found to have T_d of 61°C at pH 4.5. Five of the above variants were found to have T_d of 64-66°C, i.e. an improvement of 3-5°.

Hydrolysis of BETEB

The thermostability of the *H. insolens* cutinase and two of the above variants was measured by hydrolysis of BETEB at elevated temperature. For each cutinase, the following mixture was incubated for 17 hours at various temperatures in the range 55-70°C:

- 0.1 ml 0.5 M glycyl-glycine buffer (pH 8.5)
- 0.1 ml 0.5 % BETEB dissolved in ethanol
- 0.1 ml enzyme solution (approx. 25 LU/ml)
- 30 0.7 ml Milli Q water

The degree of hydrolysis was measured after the incubation. The results are shown in the table below.

·	Variant	Variant	Parent
	27 LU/ml	25 LU/ml	24 LU/ml
55°C	98 %	99 %	72 %
60°C	91 %	83 %	33 %
65°C	66 %	13 %	7 %

These results clearly show that the variants have improved thermostability compared to the parent cutinase.

Hydrolysis of BETEB

The thermostability of the *H. insolens* cutinase and three of the above variants was measured by hydrolysis of BETEB at 60°C for 2 hours. The hydrolysis was carried out at the above conditions, except that the temperature was fixed at 60°C and the cutinase dosage was varied. The results below are shown in the table below.

LU/ml	Variant	Variant	Variant	Parent
0	0 %	0 %	0 %	0 %
10	97 %	99 %	9 %	6 %
20	98 %	99 %	74 %	
50	98 %	94 %	93 %	15 %
100	88 %	69 %	92 %	34 %
300				41 %
600				63 %
1200				82 %

The results show a much faster hydrolysis at 60°C with the variants than with the parent cutinase.

Example 4: Hydrolysis of c3ET

The *H. insolens* cutinase and five of the above variants were tested in hydrolysis of c3ET at elevated temperature. For each cutinase, the following mixture was incubated for 2 hours at various temperatures.

5 0.115mg c3ET (0.1ml of 2mM c3ET dissolved in HFIP was taken in reaction vessel. Solvent was removed under vacuum, then dried up at 70°C over night)

0.1ml 0.5M glycyl-glycine buffer (pH8.5)

0.1ml enzyme solution (approx. 600LU/ml)

0.8ml Milli Q water

After the incubation, 2ml of 1,1,1,3,3,3-Hexafluoro-2-propanol (HFIP) was added to each reaction mixture, then hydrolysis ratio was measured by HPLC. The results shown in Fig 3 clearly indicate that the variants have improved thermostability compared to the parent cutinase.

Example 5: Hydrolysis of c3ET on yarn

The thermostability of the *H. insolens* cutinase five of the above variants was tested using polyester yarn containing c3ET as by product. The following substrate mixture was preincubated at 60 or 65°C:

0.1g polyester varn

0.2ml 0.5M glycyl-glycine buffer (pH8.5)

20 1.7ml Milli Q water

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After preincubation, 0.1ml enzyme solution (approx. 1000 LU/ml) was added to each reaction vessel and incubated for 17 hours. Then 2ml HFIP was added and left for 30 minutes to extract and hydrolyze c3ET sitting on the surface of the polyester yarn; then the hydrolysis ratio was measured. The results are shown in Fig. 4.

It is seen that the variants are more effective than the parent cutinase for hydrolyzing c3ET on polyester yarn. One variant gives higher hydrolysis ratio at 65°C than at 60°C.

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Example 6: Treatment of yarn with cutinase variant

Time courses of c3ET hydrolysis on polyester yarn at different temperature or dosage were examined. Time course at different temperatures is shown in Fig 5. It is seen that the optimum temperature is 65°C. At 70°C there is still about half of the activity left. Time course with increased enzyme dosage is shown in Fig 6. The curves at dosage 275 and 550 LU/ml are seen to be the same, indicating that the hydrolysis ratio reached to plateau between dosage of 100 to 275 LU/ml. Presumably 200LU/ml is enough.

Example 7: Dyeing polyester with reactive dye

The following polyester fabrics were treated:

woven fabric; ca. 2 x 2 cm, 34mg

knitted fabric; ca. 1.5 x 1.5 cm, 50mg

Each fabric was soaked in 0.9 ml, 50 mM GlyGly (glycyl-glycine) buffer (pH 8.5) and 0.1 ml solution of a variant of the *H. insolens* cutinase (1100 LU/ml), and incubated at 65 or 70°C. After one day, another 0.1 ml enzyme solution was added, incubation was continued for two more days, the fabrics were then taken out and rinsed in water. A comparative experiment was made with the parent cutinase, and a blank was treated in the same manner without enzyme.

The fabrics were stirred in a mixture of 9 g 120 g Na₂SO₄ and 60 g Na₂CO₃ in 3 liter deionized water at 60 °C for 30 min, and then rinsed with running warm water. The reactive dye was Celmazol Brilliant Blue B (product of Mitsui Chemical Co., Japan), which has the structure Chromophore-NHPh-SO₂CH₂CH₂OSO₃Na.

In all four experiments, (woven and knitted, 65 and 70°C), the fabrics were uniformly dyed.

25 Example 8: Solubilization of polyester fragments from knitted textile

A 1x1 cm sample of knitted polyester textile (PET, polymer of ethyleneglycol and terephthalic acid) was incubated for 1 hour in 1 ml of buffer at pH 10, 60°C with 0.01 mg of a variant of *H. insolens* cutinase. The reaction mixture was separated, and the release of terephthalic acid was found by measuring OD at 250 nm (ex-

PCT/DK99/00678 WO 00/34450

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pressed as OD₂₅₀/mg PET) comparative experiments made without were enzyme or with the parent cutinase. Results:

	Enzyme	OD ₂₅₀
Invention	Cutinase variant	4.5
Reference	Parent cutinase	0.3
	None	0.1

The results show that the variant is effective in solubilizing polyester.

In another experiment, the cutinase variant was tested for 2 hours at 65°C 5 with and without the addition of a non-ionic surfactant (alcohol ethoxylate, product name Softanol 50), using various amounts of the variant from 0.5 to 200 LU/ml. The results showed more solubilization in the presence of non-ionic surfactant.

Example 9: Hydrolysis of polycaprolactone and polyester film

About 0.1 g of polycaprolactone or polyester film were put in tubes. They were soaked in 5ml of 50mM GlyGly buffer (pH 8.5) with or without a variant of H. insolens cutinase (450 LU). They were incubated at 70°C for 5 hours. After the reaction we observed a thin layer of hydrolysate on the surface of the tubes with enzyme. both with polycaprolactone and with polyester film. On the other hand no change 15 was observed in controls without enzyme. In the case of polycaprolactone there was 10% of weight loss. We see no weight change of polyester.

Example 10: cPET hydrolysis

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The performance of a cutinase variant was compared with the parent enzyme (H. insolens cutinase). The trials were done as follows:

An oligomer-stained swatch of (black) PET-fabric (app. 4cm x 13cm) is subjected to the enzyme-treatment at relatively low agitation in a so-called minitergitometer apparatus. The PET-fabric is mounted onto a cylindrical, perforated holder (radius ca.2 cm, height ca 6 cm), that rotates around its axis, and with the oligomer stained side of the PET fabric facing the exterior of the cylinder.

The fabric is immersed in a 150ml glass-beaker containing 100ml of the treatment solution at a given temperature (here 65°C). After a given treatment time

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(here 90minutes) the PET swatch is removed from the bath and rinsed in deionized water and air dried.

After conditioning the swatches are visually ranked (with respect to oligomer stain removal) on the side having the oligomer-staining. The rating being as follows:

-2: Sample significantly worse than blank (no enzyme)

-1: Sample slightly worse than blank (no enzyme)

0: Sample can not be distinguished from blank

1: Sample slightly improved vs blank

2: Sample significantly improved over blank

Also, the swatches are read spectrofotometrically (apparatus: Hunterlab Reflectometer) to quantify the color strength (K/S-value at 600nm).

The table below summarizes the test-conditions for a trial comparing the performance the enzymes under similar conditions:

Temperature:

65°C

Buffer/pH:

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50 mM glycine buffer, pH 10.3

Treatment time (min)

90

Dosage of Enzyme (LU/g)

30000

Results from the trial are summarized below

Enzyme	Visual rating (avg.)	K/S Difference @ 600 nm
None	0 (defined)	2.33
Parent cutinase	0	2.38
Cutinase variant	1.5-2.0	2.89

From this set of experiments it thus appears that the parent enzyme provides no or only very limited effect at the given test conditions (probably because the temperature is too high for the enzyme to retain activity), while the cutinase variant provides a substantial removal of the oligomer staining from the PET-fabric.

Example 11: cPET hydrolysis

The pH and temperature profile of a variant of *H. insolens* cutinase was tested in a model disperse dyeing experiment. The trials were performed as follows:

An oligomer-stained swatch of (black) PET-fabric is subjected to the conditions of a typical disperse dyeing sequence in a Werner Mathis Labornat. In overview of the process, the swatch is added to a buffer solution, heated to 130°C,
cooled down to the treatment temperature. Enzyme or buffer is added and then held
at the desired temperature for 30 minutes. The solution is cooled down to room temperature and turbidity in the wash liquor is measured. The reduction in turbidity is a
direct measure of the cutinase activity, corresponding to hydrolyzed cPET oligomers.

Detailed description of the experiment:

A black PET (app. 4cm \times 13cm) swatch is added 140 ml 100 mM Britton-Robinson buffer containing 0.2 g/l Lutensol AT11 (BASF) and loaded in the Laborat (32 rotation per minute).

The Laborat is heated to 130°C at a gradient of 9°C/minute, and held for 10 minutes.

The beakers are cooled to run temperature (according to table below) at a gradient of 9°C/minute, and held for 1 minute.

10 mL enzyme solution (100 LU/ml of the variant) or buffer solution (0 LU/ml) 20 at appropriate pH is injected to the beakers.

The Labornat is re-heated to temperature at a gradient of 2°C/minute, and held for 30 minutes.

The swatches are removed, and the wash liquor is cooled down to room temperature.

Turbidity of the wash liquors are measured.

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Evaluation: Turbidity is measured on Hach 18900 Ratio Turbidimeter (standardized with 1.8, 18, and 180 NTU Turbidity Standards). Enzyme performance is calculated relative to a blank as the difference between turbidity of blank liquor (no enzyme) and turbidity of enzyme treated liquor.

The relative performance (reduction in turbidity) of the cutinase variant is calculated, and the results are shown in the following table. When a negative num-

27 ber is obtained, then the result is given as "negative". A negative number is assumed to be an artifact, caused by the variation of the set up.

Temperature	pH 7	pH 8	pH 9	pH 10
60°C	39	57	37	14
65°C	39	16	60	30
70°C	25	12	54	33
75°C	22	50	114	58
85°C	negative	negative	15	negative

The results show that the cutinase variant is active over a broad pH and 5 temperature range, with optimum oligomer removal in the current set up around pH 9 and 75°C. Inactivation seems to occur at or above 85°C.

Example 12: cPET hydrolysis

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The effect of treatment time was investigated for a variant of H. insolens cutinase in a model disperse dyeing experiment. The trials were performed as follows:

An oligomer-stained swatch of (black) PET-fabric is subjected to the conditions of a typical disperse dyeing sequence in a Werner Mathis Laborat. In overview of the process, the swatch is added to a buffer solution, heated to 130°C, cooled down to the treatment temperature. Enzyme or buffer (100 mM Britton-Robinson pH 9) is added, and then held at 75°C for 0-40 minutes. The solution is 15 cooled down to room temperature and turbidity in the wash liquor is measured. The reduction in turbidity is a direct measure of the cutinase activity, corresponding to hydrolyzed cPET oligomers.

Detailed description of the experiment:

A black PET (app. 4cm x 13cm) swatch is added to 140 ml 100 mM Britton-20 Robinson buffer containing 0.2 g/l Lutensol AT11 (BASF) and loaded in the Laborat (32 rotation per minute).

The Laborat is heated to 130°C at a gradient of 9°C/minute, and the temperature is held for 10 minutes.

The beakers are cooled to 75°C at a gradient of 9°C/minute, and held for 1 25 minute.

10 mL enzyme solution (100 LU/ml of variant) or 100 mM Britton-Robinson buffer pH 9.0 (0 LU/ml) is injected into the beakers.

The Laborat is re-heated to 75°C at a gradient of 2°C/minute, and held for the appropriate number of minutes (0-40 minutes, see table below).

The swatches are removed, and the wash liquor is cooled down to room temperature.

Turbidity of the wash liquors are measured.

Evaluation: Turbidity is measured on Hach 18900 Ratio Turbidimeter (standardized with 1.8, 18, and 180 NTU Turbidity Standards). Enzyme performance is calculated relative to a blank at time equal to zero: Turbidity of blank liquor at time zero (no enzyme) subtracted turbidity of enzyme treated liquor (at a given time).

The relative performance (reduction in turbidity) of the cutinase variant was calculated, and the results are shown in the following table.

Time (minutes)	Relative perform- ance (Reduction in turbidity)
0	0
5	42
10	48
15	62
20	69
25	85
30	72
40	78

The results show that the effect of the enzyme is increased over time. At the current enzyme dose and oligomer concentration, it seems to level off above approx. 20 minutes.

Example 13: Fiber modification

The effect on wetting characteristics of a disperse dyed polyester fabric was investigated by treating the fabric with a variant of *H. insolens* cutinase prior to dye-

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ing. The experiment therefore consisted of two phases, the actual fiber modification and the disperse dyeing pro- cedure.

Phase 1 - Fiber Modification:

Equipment:

Atlas Launder-O-meter LP2

Fabric:

knit 100 % scoured polyester from Testfabrics

:Hq

50 mM potassium phosphate buffer, pH 7

Abrasives:

5 big steel balls

Beaker Vol.:

120 mL

Treatment:

2 hours 65°C then ramped up to 90°C and held for 1 hour

Swatch Prep:

Cut 3* 1.5 g swatch of fabric, 3 per beaker = 4.5 g

Rinse:

5 Rinse in deionized water.

Phase 2 - Dyeing - disperse dye:

Dye Solution:

Add together with deionized water to make liquor ratio 1:20-

0.4 % Dianix Red (DyStar) SE-CB (owf)

10 pH to 4.5 - 5

Dyeing Procedure:

- 1. One swatch per treatment from the fiber modification is used for the dyeing (1.5 g/swatch is used for the liquor ratio calculation).
- Make dyebath according to the recipe above. Add the cold dye solution
 to the Laborat beakers and heat to 55°C at a gradient of 3.5°C/minute. Run for 5 minutes once temperature has been reached.
 - 3. Add the fabric to the beaker.
 - 4. Raise temperature to 130°C at a gradient of 1.5°C/minute. Dye for 30 minutes.
- 5. Cool to 70°C at a gradient of 5°C/minute. Drop bath, but collect, and rinse fabric hot (60°C) for 10 minutes. Follow the hot rinse with a room temperature overflow rinse until all bleeding had stopped.
 - 6. Let air dry overnight.

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Tests/Analysis:

AATCC Test Method 61 - Colorfastness to washing

Percent Dyebath Exhaustion - Spectrophotometer

K/S and L* - Reflectometer

5 AATCC TM-79 Drop Test

Results:

The results from the fiber modification are shown in the following table.

Variant dosage	Staining (AATCC TM- 61)	Color Change (K/S @ 530 before and after TM-61)	Drop Test (AATCC TM-79)
Blank	4.5	5	53 sec.
50 LU/mL	4.5	5	18 sec.
100 LU/mL	4.5	5	15 sec.

The results show that the treatment of polyester with the variant increases the wetting substantially. No adverse effects are noticed on the dyeability with the disperse dye in the current set-up.

Example 14: Malodor reduction in textiles soiled with human sweat/sebum by use of a cutinase variant in laundry

The performance of cutinase, with respect to malodor reduction, can be tested in a one-cycle washing trial carried out in a Terg-O-tometer.

Experimental conditions:

Washing liquor: 1000 ml per beaker

Swatches: 100 % polyester (interlock knitted, previously cleaned by Soxhlet extraction). 24 swatches (3.3 × 3.5 cm) per beaker.

Soil: Human male axillary sweat and sebum applied by wiping the armpits after exercise.

Detergent: 5 g/L of a standard color detergent. No pH adjustment.

Water hardness: 3.2 mM Ca²⁺/Mg²⁺ (in a ratio of 5:1)

Wash Temperature: 30°C

25 Wash time: 30 min

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Rinse: 15 minutes in running tap water

Evaluation:

After wash the wet swatches are placed in closed, tinted 200 ml glasses. A trained sensory panel (9-11 judges) evaluates the odor by sniffing the headspace over the wet samples and evaluates the total odor intensity. The odor intensity is noted by placing a mark on an unstructured line scale measuring 15 cm, with word anchors at each end ('nothing' at the beginning of the scale and 'very strong' at the end). All evaluations are performed twice. The swatches are evaluated on day 1, 2 and 3 after wash (swatches are kept in the glasses at all times).

CLAIMS

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- 1. A variant of a parent fungal cutinase, which variant:
 - comprises substitution of one or more amino acid residues at a position which is located:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid, and
 - b) is more thermostable than the parent cutinase.
- 10 2. The variant of the preceding claim which comprises substitution of one or more amino acid residues at a position which is located:
 - i) within 12 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 15 positions from the N-terminal amino acid.
 - 3. A variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which is located:
 - a) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 20 positions from the N-terminal amino acid,

with the proviso that it is not a variant of the cutinase of *Fusarium solani pisi* having one of the substitutions R17, T18, T19V, D21N, I24E, Y38F, R40, G41A, S42, T43, E44, T45, G46, N47R, G49, T50, L51, P53, S54, A56C, S57, N58R, S61, A62E, K65A, D66S, G67D, W69Y, I70C, G74, G75, R78, Y119, G192, P193, D194R, 25 A195, R196, G197V, or A199C (*Fusarium solani pisi* cutinase numbering).

- 4. A variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which:
 - a) has a solvent accessible surface, and

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- b) is located:
 - i) within 17 Å from the location of the Nterminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid,

with the proviso that it is not a variant of the cutinase of *Fusarium solani pisi* having one of the substitutions T18, Y38F, R40, G41A, S42, T43, E44, T45, N47R, G49, T50, L51, P53, S54, A56C, A62E or G192 (*Fusarium solani pisi* cutinase numbering).

- 10 5. A variant of a parent fungal cutinase comprising substitution of one or more amino acid residues which is located:
 - a) less than 12 Å from the location of the N-terminal amino group (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 15 positions from the N-terminal amino acid.
- with the proviso that the variant is not the cutinase of *Fusarium solani pisi* having one of the substitutions R17, T18, T19V, D21N, Y38F, R40, T45, G46, N47R, G49, T50, L51, P53, S54, A56C, S57, N58R, K65A or I70C (*Fusarium solani pisi* cutinase numbering).
- 6. The variant of any preceding claim wherein the parent cutinase is native to a filamentous fungus, preferably a strain of *Humicola* or *Fusarium*, preferably *H. insolens* or *F. solani pisi*, most preferably *H. insolens* strain DSM 1800.
 - 7. The variant of any preceding claim wherein the parent cutinase has an amino acid sequence which can be aligned with the cutinase of *H. insolens* strain DSM 1800.
- 25 8. The variant of any preceding claim wherein the parent cutinase has an amino acid sequence which is at least 50 % homologous to the cutinase of *H. insolens* strain DSM 1800, preferably at least 70 % homologous, more preferably at least 80 % homologous.

- 9. A variant of a parent fungal cutinase from *Humicola insolens* which comprises substitution of one or more amino acid residues located:
 - a) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 20 positions from the N-terminal amino acid.

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- 10. The variant of the preceding claim which comprises substitution of one or more amino acid residues located:
 - a) less than 12 Å from the location of the N-terminal amino group (as calculated from amino acid residues in a crystal structure), and/or
 - b) within 15 positions from the N-terminal amino acid
- 11. The variant of any preceding claim which comprises substitution of one or more amino acids having a solvent accessible surface.
- 12. The variant of any preceding claim wherein one or more substitutions is substitution of a negatively charged amino acid with a neutral or positively charged amino acid or substitution of a neutral amino acid with a positively charged amino acid.
- 13. The variant of the preceding claim wherein one or more substitutions is at a position corresponding to position E6, E10, E30, E47, D63, E82 and/or E179 in the cutinase of *Humicola insolens* strain DSM 1800, preferably a substitution with 20 R/K/Y/H/Q/N, more preferably a substitution corresponding to E6N/Q, E10N/Q, E47K/R and/or E179N/Q (*H. insolens* cutinase numbering).
 - 14. The variant of any preceding claim wherein one or more substitutions is substitution with a Pro residue, preferably at a position corresponding to position A14 and/or R51.
- 25 15. The variant of any preceding claim which has one, two, three, four, five or six of said substitutions.

- 16. The variant of any preceding claim which has substitutions corresponding to one of the following in the cutinase of *Humicola insolens* strain DSM 1800:
 - a) R51P
 - b) E6N/Q + L138I
- 5 c) A14P + E47K
 - d) E47K
 - e) E179N/Q
 - f) E6N/Q + E47K + R51P
 - g) A14P + E47K + E179N/Q
- 10 h) E47K + E179N/Q
 - i) E47K + D63N
 - i) E6N/Q + A14P + E47K + R51P + E179N/Q
 - k) E6N/Q + E10N/Q + A14P + E47K + R51P + E179N/Q, or
 - l) Q1P + L2V + S11C + N15T + F24Y + L46I + E47K
- 15 17. The variant of any preceding claim which has hydrolytic activity towards terephthalic acid esters, particularly towards cyclic tri(ethylene terephthalate) and/or Terephthalic acid bis(2-hydroxyethyl)ester dibenzoate (BETEB).
 - 18. The variant of any preceding claim which has a denaturation temperature which is at least 5° higher than the parent cutinase, preferably measured at pH 8.5
- 20 19. A DNA sequence encoding the variant of any preceding claim.
 - 20. A vector comprising the DNA sequence of the preceding claim.
 - 21. A transformed host cell harboring the DNA sequence of claim 19 or the vector of claim 20.
 - 22. A method of producing the variant of any of claims 1-18 comprising
- 25 a) cultivating the cell of claim 21 so as to express and preferably secrete the variant, and

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b) recovering the variant.

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- 23. A method of constructing a cutinase variant, which method comprises:
 - a) selecting a parent fungal cutinase,
- 5 b) identifying one or more amino acid residues in the parent cutinase at positions which are:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid, and
 - c) making alterations each of which is an insertion, a deletion or a substitution of the amino acid residue,
 - optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),
 - e) preparing the variant resulting from steps b-d,
 - f) testing the thermostability of the variant,
 - g) optionally repeating steps b-f, and
- h) selecting a variant having higher thermostability than the parent cuti nase.
 - A method of producing a cutinase variant, which method comprises:
 - a) selecting a parent fungal cutinase,
 - b) identifying one or more amino acid residues in the parent cutinase at positions which are:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid, and
- c) making alterations each of which is an insertion, a deletion or a substitution of the amino acid residue,

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d) optionally, making alterations each of which is an insertion, a deletion or a substitution of an amino acid residue at one or more positions other than b),

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- e) preparing the variant resulting from steps b-d,
- f) testing the thermostability of the variant,
 - g) optionally repeating steps b-f,
 - h) selecting a variant having higher thermostability than the parent cutinase, and
 - i) producing the variant to obtain the cutinase variant.
- 10 25. A process for enzymatic hydrolysis of a cyclic oligomer of poly(ethylene terephthalate), which process comprises treating the cyclic oligomer with a variant of a parent fungal cutinase, which variant comprises substitution of one or more amino acid residues at a position which is located:
 - i) within 17 Å from the location of the N-terminal amino acid (as calculated from amino acid residues in a crystal structure), and/or
 - ii) within 20 positions from the N-terminal amino acid.
 - 26. The process of the preceding claim, in which the cyclic oligomer is cyclic tri(ethylene terephthalate).
- 20 27. The process of claim 25 or 26 wherein the treatment is done at 60-80°C, preferably at 65-75°C.
 - 28. The process of any of claims 25-27 wherein the cyclic oligomer is present in and on the fibers of a polyester containing fabric or yarn.
- 29. The process of any of claims 25-28 which further comprises subsequently rinsing the fabric or yarn, preferably rinsing with an aqueous solution having a pH in the range of from about pH 7 to about pH 11.
 - 30. A process for dyeing polyester fabric or yarn, comprising:

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a) treating the fabric or yarn with a cutinase having a thermal denaturation temperature of 65°C or higher at pH 8.5; and

- b) dyeing the treated fabric with a reactive dye or a disperse dye.
- 5 31. The process of the preceding claim wherein the cutinase is the variant of any of claims 1-18.
 - 32. A detergent composition comprising a surfactant and the variant of any of claims 1-18.
- 33. A method for detecting cutinase activity in a sample, comprising incubating the sample with terephthalic acid bis(2-hydroxyethyl)ester dibenzoate and detecting hydrolysis of said ester.
 - 34. A process for improving the functional finish of a PET-containing yarn or fabric comprising
 - a) treating the yarn or fabric with the variant of any of claims 1-18, and
- b) subsequently the yarn or fabric with a finishing agent selected from the group consisting of softeners, anti-crease resins, anti-static agents, anti-soiling agents.

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Fig. 1
3D structure of cutinase from *Humicola insolens*

ATOM	1	N	GLY	Δ	3	24.42	4 -7.935	18.390	1.00 46.73
ATOM	2	CA	GLY		3	23.84		17.546	1.00 42.29
ATOM	3	C	GLY		3		6 -10.112	16.727	1.00 37.35
	4	0	GLY		3	25.34		16.728	1.00 37.33
ATOM	5	N	ALA		4	23.66		15.728	1.00 33.38
ATOM									
ATOM	6	CA	ALA		4	23.05		14.555	1.00 30.95
ATOM	7	С	ALA		4		4 -11.246	14.920	1.00 28.33
ATOM	8	0	ALA		4		7 -10.499	14.446	1.00 22.94
MOTA	9	CB	ALA		4		4 -11.780	13.556	1.00 26.92
MOTA	10	N	ILE		5	21.58		16.043	1.00 26.48
MOTA	11	CA	ILE		5	20.28		16.637	1.00 25.65
MOTA	12	С	ILE	Α	5	20.31	6 -12.151	18.118	1.00 22.40
ATOM	13	0	ILE	A	5	21.06	0 -12.888	18.717	1.00 24.74
ATOM	14	CB	ILE	Α	5	19.72	4 -13.683	16.524	1.00 26.04
ATOM	15	CG1	ILE	Α	5	19.85	2 -13.927	15.050	1.00 29.85
ATOM	16	CG2	ILE	Α	5	18.37	4 -13.558	17.159	1.00 20.48
ATOM	17	CD1	ILE	Α	5	19.06	6 -15.133	14.709	1.00 27.96
ATOM	18	N	GLU	Α	6	19.46	1 -11.377	18.668	1.00 20.52
ATOM	19	CA	GLU	A	6		7 -11.015	20.040	1.00 17.94
ATOM	20	С	GLU		6	17.71		20.432	1.00 17.76
ATOM	21	Ō	GLU		6	16.93		19.990	1.00 17.60
ATOM	22	CB	GLU		6	19.80		20.199	1.00 14.22
ATOM	23	CG	GLU		6	21.23		20.385	1.00 16.71
ATOM	24	CD	GLU		6		8 -10.387	21.030	1.00 34.47
ATOM	25	OE1	GLU		6	21.63		21.693	1.00 34.47
ATOM	26	OE2	GLU		6		0 -10.310	20.975	1.00 43.37
ATOM	27	N	ASN		7		5 -11.895	21.333	1.00 37.43
		CA	ASN		7				
MOTA	28				7	16.07		21.846	1.00 24.04
ATOM	29	C	ASN			15.92		23.238	1.00 22.08
ATOM	30	0	ASN		7	15.09		23.820	1.00 24.00
ATOM	31	CB	ASN		7		8 -13.307	21.820	1.00 25.06
MOTA	32	CG	ASN		7	15.03		20.341	1.00 38.52
MOTA	33	OD1			7	15.51		19.759	1.00 48.45
ATOM	34		ASN		7	14.31		19.968	1.00 36.89
MOTA	35	N	GLY		8	16.67		23.926	1.00 23.56
MOTA	36	CA	GLY		8	16.65	4 -10.628	25.363	1.00 23.69
ATOM	37	С	GLY	Α	8	15.36	6 - 10.247	25.984	1.00 22.72
MOTA	38	0	GLY	Α	8	14.96	7 -10.939	26.867	1.00 32.25
MOTA	39	N	LEU	Α	9	14.78	5 -9.144	25.755	1.00 23.61
ATOM	40	CA	LEU	Α	9	13.47	0 -8.753	26.033	1.00 23.73
ATOM	41	С	LEU	Α	9	12.55		25.782	1.00 25.93
ATOM	42	0	LEU		9	11.49		26.480	1.00 30.47
ATOM	43	СВ	LEU		9	12.97		25.105	1.00 5.84
ATOM	44	CG	LEU		9	11.55		25.470	1.00 23.25
ATOM	45		LEU		9	11.42		26.968	1.00 20.21
ATOM	46		LEU		9	11.00		24.714	1.00 20.21
ATOM	47	N	GLU		10		5 -10.786	24.773	1.00 17.04
ALI OF	3 /	14	010	17	10	12.11	J TO: 100	27.113	1.00 29.30

7.5004	40	C7	CTIT	70	10	11 635	-11.681	24.484	1.00	33.93
ATOM	48 49	CA C	GLU GLU		10		-12.872	25.412		32.18
ATOM			GLU		10		-13.159	25.996		36.67
ATOM	50	0					-11.996	23.012		40.97
MOTA	51	CB	GLU		10		-12.303	22.745		51.96
ATOM	52	CG	GLU		10			21.437	1.00	54.08
ATOM	53	CD	GLU		10		-11.711	20.635	1.00	48.22
MOTA	54	OE1	GLU		10		-11.440	21.471		52.39
ATOM	55	OE2	GLU		10		-11.643			29.58
ATOM	56	N	SER		11		-13.334	25.688		
ATOM	57	CA	SER		11		-14.455	26.645		35.25
ATOM	58	C	SER		11		-14.012	28.047		39.86
ATOM	59	0	SER		11	13.688	-14.790	28.919		43.72
ATOM	60	CB	SER		11.		-15.364	25.983		33.73
ATOM	61	OG	SER	Α	11	15.275	-14.620	25.928		46.98
ATOM	62	N	\mathtt{GLY}		12 .	13.467	-12.802	28.456		41.40
ATOM	63	CA	GLY	Α	12	13.841	-12.332	29.752		45.34
ATOM	64	С	\mathtt{GLY}	Α	12		-12.562	30.694		47.62
ATOM	65	0	\mathtt{GLY}	Α	12	11.485	-12.335	30.335	1.00	50.76
ATOM	66	N	SER	Α	13		-12.900	31.936		48.09
ATOM	67	CA	SER	Α	13		-13.158	32.995		45.26
ATOM	68	С	SER	A	13		-11.933	33.772	1.00	39.53
ATOM	69	0	SER	Α	13	12.563	-11.204	33.992		36.30
ATOM	70	CB	SER	Α	13	12.708	-14.006	34.101	1.00	51.20
ATOM	71	OG	SER	Α	13		-13.947	35.338		57.14
ATOM	72	N	ALA	A	14	10.256	-11.785	34.214	1.00	35.22
ATOM	73	CA	ALA	Α	14	10.068	-10.530	34.964	1.00	34.78
ATOM	74	С	ALA	Α	14	10.574	-10.620	36.417	1.00	37.51
MOTA	75	0	ALA	Α	14	10.809	-9.584	37.113	1.00	38.41
ATOM	76	СВ	ALA	A	14	8.714	-9.915	34.903	1.00	32.71
ATOM	77	N	ASN	Α	15	11.039	-11.834	36.737	1.00	38.85
ATOM	78	CA	ASN	A	15	11.715	-12.086	37.963	1.00	
ATOM	79	С	ASN	A	15	13.073	-11.411	37.953	1.00	46.45
ATOM	80	Ō	ASN		15	13.453	-11.022	39.022	1.00	52.50
ATOM	81	СВ	ASN		15	12.088	-13.533	38.207	1.00	53.08
ATOM	82	CG	ASN		15	10.772	-14.226	38.553	1.00	71.86
ATOM	83		ASN		15	9.837	-13.535	38.998	1.00	71.73
ATOM	84		ASN		15	10.866	-15.523	38.267	1.00	77.71
ATOM	85	N	ALA		16		-11.305	36.812	1.00	46.73
ATOM	86	CA	ALA		16		-10.470	36.743	1.00	41.22
ATOM	87	C	ALA		16	15.031	-9.286	35.798	1.00	36.70
ATOM	88	0	ALA		16	16.027		35.075	1.00	37.67
ATOM	89	CB	ALA		16		-11.545	36.301	1.00	41.80
ATOM	90	N	CYS		17	14.300	-8.227	35.843	1.00	30.62
ATOM	91	CA	CYS		17	14.614		34.997		31.78
ATOM	92	C	CYS		17	16.024		35.149		32.94
ATOM	93	0	CYS		17	16.744		36.113		39.10
	94	CB	CYS		17	13.679		35.138		28.00
ATOM		SG	CYS		17	12.048		34.858		24.72
ATOM	95	36	CIC	, 7.	± '	12.040	0.005	5		

			220	_				24 222	
ATOM	96	N	PRO		18	16.529	-5.910	34.092	1.00 30.49
ATOM	97	CA	PRO		18	17.994	-5.626	33.971	1.00 22.04
ATOM	98	С	PRO		18	18.178	-4.138	34.241	1.00 20.15
ATOM	99	0	PRO		18	17.085	-3.459	34.370	1.00 17.83
ATOM	100	CB	PRO		18	18.353	-6.003	32.559	1.00 19.20
ATOM	101	CG	PRO		18	17.044	-6.595	32.101	1.00 20.16
ATOM	102	CD	PRO		18	15.903	-5.936	32.792	1.00 24.35
ATOM	103	N	ASP		19	19.428	-3.652	34.011	1.00 14.85
ATOM	104	CA	ASP	Α	19	19.451	-2.168	34.226	1.00 16.59
ATOM	105	C	ASP	Α	19	18.739	-1.367	33.156	1.00 20.42
ATOM	106	0	ASP	Α	19	18.311	-0.242	33.430	1.00 23.84
ATOM	107	CB	ASP	Α	19	20.896	-1.818	34.485	1.00 27.25
ATOM	108	CG	ASP	Α	19 ·	21.433	-2.389	35.793	1.00 42.30
ATOM	109	OD1	ASP	A	19	21.162	-3.549	36.297	1.00 53.52
ATOM	110	OD2	ASP		19	22.251	-1.719	36.543	1.00 54.02
ATOM	111	N	ALA		20	18.646	-1.780	31.895	1.00 20.18
ATOM	112	CA	ALA		20	18.066	-1.036	30.809	1.00 17.43
ATOM	113	C	ALA		20	17.713	-2.087	29.703	1.00 16.06
ATOM	114	Ö	ALA		20	18.334	-3.172	29.860	1.00 9.45
ATOM	115	CB	ALA		20	18.975	-0.048	30.100	1.00 12.07
ATOM	116	N	ILE		21	16.814	-1.602	28.829	1.00 12.07
ATOM	117	CA		Α	21	16.657	-2.583	27.753	1.00 9.23
ATOM	118	C		A	21	16.952	-1.745	26.486	1.00 3.23
ATOM	119	0		A	21	16.681	-0.473	26.403	1.00 14.77
ATOM	120	CB		A	21	15.208	-2.984	27.837	1.00 12.01
ATOM	121	CG1	ILE		21				
ATOM	122	CG2				14.851	-3.898	28.956	1.00 15.55
			ILE		21	14.689	-3.671	26.514	1.00 13.71
ATOM	123	CD1	ILE		21	13.401	-3.879	29.372	1.00 6.12
ATOM	124	N	LEU		22	17.432	-2.451	25.391	1.00 12.24
ATOM	125	CA	LEU		22	17.665	-1.774	24.087	1.00 11.27
ATOM	126	C	LEU		22	16.849	-2.517	23.038	1.00 14.60
ATOM	127	0	LEU		22	16.908	-3.781	22.850	1.00 9.78
ATOM	128	CB	LEU		22	19.087	-1.865	23.693	1.00 10.96
ATOM	129	CG	LEU		22	19.493	-1.543	22.257	1.00 10.32
ATOM	130	CD1	LEU		22	19.311	-0.081	21.900	1.00 4.72
ATOM	131	CD2	LEU	Α	22	20.990	-1.842	22.156	1.00 7.42
ATOM	132	N	ILE	Α	23	16.038	-1.815	22.242	1.00 15.13
ATOM	133	CA	ILE	Α	23	15.298	-2.459	21.115	1.00 18.06
ATOM	134	С	ILE	Α	23	15.916	-1.771	19.901	1.00 17.42
ATOM	135	0	ILE	A	23	16.117	-0.519	19.795	1.00 19.31
ATOM	136	CB	ILE	Α	23	13.820	-2.194	21.392	1.00 18.16
ATOM	137	CG1	ILE		23	13.208	-3.076	22.447	1.00 14.23
ATOM	138	CG2	ILE		23	12.787	-2.167	20.247	1.00 13.19
ATOM	139	CD1	ILE		23	12.142	-2.065	22.976	1.00 20.41
ATOM	140	N	PHE		24	16.218	-2.548	18.940	1.00 14.59
ATOM	141	CA	PHE		24	16.859	-2.159	17.671	1.00 11.72
ATOM	142	C	PHE		24	16.347	-2.719	16.353	1.00 7.25
ATOM	143	0	PHE		24	16.095	-3.998	16.161	1.00 7.23
 -	~	-				10.000	J. J. J. U	TO - TO T	1.00 3.47

ATOM	144	CB	PHE		24	18.195	-2.855	17.658		12.61
ATOM	145	CG	PHE		24	19.015	-2.150	16.716		10.72
ATOM	146	CD1			24	19.457	-0.844	16.913		13.08
ATOM	147		PHE		24	19.325	-2.852	15.558	1.00	6.61
ATOM	148	CE1	PHE		24	20.232	-0.187	15.983	1.00	4.86
ATOM	149	CE2	PHE		24	20.061	-2.218	14.545	1.00	7.61
ATOM	150	CZ	PHE	Α	24	20.550	-0.823	14.804	1.00	8.78
MOTA	151	N	ALA	Α	25	16.037	-1.700	15.449	1.00	6.32
ATOM	152	CA	ALA	Α	25	15.662	-2.158	14.068	1.00	7.18
ATOM	153	С	ALA	A	25	16.851	-1.976	13.055	1.00	8.59
ATOM	154	0	ALA	A	25	17.518	-1.000	13.133	1.00	5.95
ATOM	155	СB	ALA	Α	25	14.488	-1.402	13.562	1.00	8.27
ATOM	156	N	ARG	Α	26	17.174	-3.032	12.325	1.00	8.84
ATOM	157	CA	ARG	A	26	18.134	-3.278	11.277	1.00	4.04
ATOM	158	С	ARG		26	17.691	-2.694	9.894	1.00	7.67
ATOM	159	0	ARG		26	16.527	-2.361	9.525	1.00	9.36
ATOM	160	CB	ARG		26	18.581	-4.659	10.756	1.00	6.06
ATOM	161	CG	ARG		26	17.705	-5.741	10.439	1.00	5.08
ATOM	162	CD	ARG		26	18.069	-7.224	10.382	1.00	6.73
ATOM	163	NE	ARG		26	17.000	-8.053	9.708	1.00	9.04
ATOM	164	CZ	ARG		26	15.724	-8.206	9.912	1.00	7.06
ATOM	165	NH1	ARG		26	15.085	-7.535	10.895	1.00	22.93
ATOM	166	NH2	ARG		26	14.809	-8.825	9.346	1.00	7.89
ATOM	167	N	GLY		27	18.761	-2.539	9.092	1.00	7.71
ATOM	168	CA	GLY		27	18.537	-1.888	7.782	1.00	5.34
ATOM	169	C	GLY		27	18.063	-2.896	6.862	1.00	4.70
ATOM	170	Õ	GLY		27	18.155	-4.139	7.075		13.14
ATOM	171	N	SER		28	17.562	-2.612	5.765		11.82
ATOM	172	CA	SER		28	17.108	-3.325	4.615		14.72
ATOM	173	C	SER		28	18.214	-4.327	4.142	1.00	7.74
ATOM	174	0	SER		28	19.286	-3.973	4.083	1.00	6.71
ATOM	175	CB	SER		28	16.460				
ATOM	176	OG	SER		28		-2.352	3.538	1.00	6.38
ATOM	177	N	THR		29	16.819	-0.978	3.833		28.10
		CA				17.942	-5.634	4.241	1.00	4.79
ATOM	178		THR		29	18.562	-6.763	3.914	1.00	8.71
ATOM	179	C	THR		29	19.500	-7.271	4.985		14.00
ATOM	180	0	THR		29	20.162	-8.326	4.713		17.68
ATOM	181	CB	THR		29	19.454	-6.680	2.617		14.90
ATOM	182	OG1			29	20.736	-6.066	2.595		14.00
ATOM	183	CG2	THR		29	18.785	-5.888	1.561		15.59
ATOM	184	N	GLU		30	19.740	-6.599	6.105		14.52
ATOM	185	CA	GLU		30	20.677	-7.266	7.056		14.10
ATOM	186	С	GLU		30	20.092	-8.513	7.647		13.07
ATOM	187	0	GLU		30	18.916	-8.726	7.705		19.98
ATOM	188	СВ	GLU		30	21.228	-6.371	8.072		15.45
ATOM	189	CG	GLU		30	21.166	-4.945	7.709	1.00	
ATOM	190	CD	GLU		30	22.073	-4.143	8.637		23.08
ATOM	191	OE 1	GLU	A	30	21.395	-3.328	9.284	1.00	19.26

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ATOM	192		GLU		30		23.317		8.712		19.71
ATOM	193	N	PRO		31		20.875		7.918		13.09
ATOM	194	CA	PRO		31			-10.818	8.402	1.00	
ATOM	195	С	PRO		31		20.167	-10.698	9.895	1.00	
ATOM	196	0	PRO		31		20.148	-9.636	10.392	1.00	20.45
ATOM	197	CB	PRO		31		21.690	-11.692	8.215	1.00	10.95
ATOM	198	CG	PRO	Α	31		22.790	-10.664	8.455	1.00	11.24
ATOM	199	CD	PRO	Α	31		22.350	-9.316	7.864	1.00	13.71
ATOM	200	N	GLY	Α	32		19.612	-11.689	10.472	1.00	18.99
ATOM	201	CA	GLY	Α	32		19.205	-11.774	11.816	1.00	13.53
ATOM	202	С	GLY	Α	32	·	18.133	-10.808	12.188		
ATOM	203	0	GLY	Α	32		17.345	-10.294	11.411	1.00	
ATOM	204	N	ASN	Α	33			-10.528	13.468	1.00	
ATOM	205	CA	ASN		33		17.290	-9.346	13.823	1.00	
ATOM	206	C	ASN		33		18.294	-8.273	14.230	1.00	
ATOM	207	Ō	ASN		33		17.774	-7.184	14.575	1.00	
ATOM	208	CB	ASN		33		16.241	-9.663	14.867		17.42
ATOM	209	CG	ASN		33						
ATOM	210		ASN				16.827	-10.201	16.127		17.97
ATOM		ND2			33		16.112	-10.395	17.089		19.05
	211		ASN		33		18.074	-10.460	16.112		13.29
ATOM	212	N		A	34		19.633	-8.378	14.282		14.22
ATOM	213	CA		A	34		20.282	-7.171	14.751		12.97
ATOM	214	C		A	34		21.142	-6.663	13.611		19.02
ATOM	215	0		A	34		21.654	-5.512	13.713		26.04
ATOM	216	CB		A	34		21.202	-7.329	15.859		13.39
ATOM	217	CG		A	34		20.579	-7.713	17.163	1.00	9.02
ATOM	218	SD	MET	Α	34	:	20.175	-6.316	18.069	1.00	9.13
ATOM	219	CE	MET	Α	34	:	21.481	-5.121	18.095	1.00	4.11
ATOM	220	N	\mathtt{GLY}	Α	35	;	21.259	-7.446	12.550	1.00	19.99
ATOM	221	CA	GLY	Α	35		22.071	-7.135	11.418	1.00	14.30
ATOM	222	С	GLY	Α	35		23.511	-7.340	11.764		17.58
ATOM	223	0	GLY	Α	35		23.965	-7.724	12.842		12.78
ATOM	224	N	ILE		36		24.450	-6.839	10.950		20.63
ATOM	225	CA	ILE		36		25.833	-7.029	11.277		17.71
ATOM	226	С	ILE		36		26.609	-5.714	11.280		16.15
ATOM	227	Ō	ILE		36		27.865	-5.618	11.662		20.30
ATOM	228	CB	ILE		36		26.412	-8.070	10.327		30.19
ATOM	229		ILE		36		26.088	-7.448	8.959		31.16
ATOM	230		ILE		36		25.944				
	231		ILE					-9.490	10.543		15.68
ATOM					36		26.922	-8.149	7.958		34.10
ATOM	232	N	THR		37		25.905	-4.589	11.040		13.00
ATOM	233	CA	THR		37		26.825	-3.396	11.141	1.00	
ATOM	234	C	THR		37		26.587	-2.513	12.350		15.44
ATOM	235	0	THR		37		27.040	-3.055	13.410		20.20
ATOM	236	CB	THR		37		26.592	-2.679	9.818		14.13
ATOM	237	OG1	THR		37		25.241	-2.212	9.503	1.00	22.62
ATOM	238	CG2	THR		37		26.949	-3.739	8.800	1.00	2.29
ATOM	239	N	VAL	A	38	;	25.733	-1.493	12.249	1.00	11.92

ATOM	240	CA	VAL		38	25.237	-0.800	13.411	1.00 15.22
ATOM	241	С	VAL		38	24.588	-1.455	14.612	1.00 14.68
ATOM	242	0	VAL	Α	38	24.906	-1.185	15.733	1.00 15.89
ATOM	243	CB	VAL	A	38	24.124	0.180	12.855	1.00 14.13
ATOM	244	CG1	VAL	A	38	23.663	0.897	14.167	1.00 13.55
ATOM	245	CG2	VAL	Α	38	24.570	1.025	11.670	1.00 6.75
ATOM	246	N	GLY	Α	39	23.745	-2.410	14.677	1.00 14.24
ATOM	247	CA	GLY	A	39	23.135	-3.151	15.746	1.00 11.03
ATOM	248	С	GLY	Α	39	24.096	-3.586	16.791	1.00 13.34
ATOM	249	0	GLY	A	39	24.131	-3.181	17.934	1.00 15.13
ATOM	250	N	PRO	A	40	25.067	-4.340	16.352	1.00 14.70
ATOM	251	CA	PRO		40 .	26.094	-5.025	17.171	1.00 13.44
ATOM	252	С	PRO		40	27.010	-3.909	17.589	1.00 11.81
ATOM	253	0	PRO		40	27.346	-3.871	18.764	1.00 12.79
ATOM	254	CB	PRO		40	26.723	-6.111	16.279	1.00 8.43
ATOM	255	CG	PRO		40	25.873	-6.243	14.950	1.00 4.84
ATOM	256	CD	PRO		40	25.198	-4.902	14.995	1.00 12.36
ATOM	257	N	ALA		41	27.226	-2.979	16.695	1.00 7.41
ATOM	258	CA	ALA		41	28.066	-1.962	17.278	1.00 11.03
ATOM	259	C	ALA		41	27.378	-1.206	18.439	1.00 14.87
ATOM	260	ō	ALA		41	28.028	-0.503	19.274	1.00 14.26
ATOM	261	CB	ALA		41	28.579	-0.905	16.313	1.00 7.17
ATOM	262	N	LEU		42	26.135	-0.811	18.237	1.00 11.87
ATOM	263	CA	LEU		42	25.487	-0.048	19.300	1.00 12.36
ATOM	264	C	LEU		42	25.337	-0.856	20.624	1.00 11.94
ATOM	265	0	LEU		42	25.423	-0.397	21.730	1.00 8.33
ATOM	266	СВ	LEU		42	24.036	0.168	18.811	1.00 13.24
ATOM	267	CG	LEU		42	23.272	1.160	19.676	1.00 15.24
ATOM	268	CD1	LEU		42	24.108	2.419	19.962	1.00 6.62
ATOM	269	CD2	LEU		42	21.991	1.580	18.943	
ATOM	270	N	ALA		43	24.905	-2.095	20.482	
ATOM	271	CA	ALA		43	24.761			1.00 10.88
ATOM	272	C	ALA		43		-3.027	21.553	1.00 12.37
ATOM	273	0	ALA		43	26.106	-3.136	22.252	1.00 15.45
ATOM	274	CB	ALA		43	25.958	-2.743	23.433	1.00 20.80
ATOM	275	N	ASN			24.148	-4.324	21.002	1.00 9.60
	276	CA	ASN		44 44	27.263	-3.440	21.636	1.00 16.91
ATOM						28.454	-3.434	22.439	1.00 20.33
ATOM	277	C	ASN		44	28.717	-2.044	23.113	1.00 17.66
ATOM	278	O	ASN		44	29.019	-1.991	24.301	1.00 17.06
ATOM	279	CB	ASN		44	29.756	-3.695	21.625	1.00 35.48
ATOM	280	CG	ASN		44	29.564	-5.115	21.138	1.00 58.23
ATOM	281		ASN		44	30.013	-5.403	20.034	1.00 79.77
ATOM	282		ASN		44	28.908	-5.945	21.921	1.00 70.10
ATOM	283	N	GLY		45	28.682	-0.988	22.297	1.00 14.39
ATOM	284	CA	GLY		45	29.015	0.221	22.976	1.00 11.65
ATOM	285	C	GLY		45	28.175	0.255	24.234	1.00 14.30
ATOM	286	0	GLY		45	28.529	0.582	25.385	1.00 10.77
ATOM	287	N	LEU	A	46	26.861	0.099	24.065	1.00 16.88

ATOM		CA LEU	A 46	25.968	0.248	3 25.20°	7 1 00 10 1
ATOM		C LEU		26.395			
ATOM		O LEU	A 46	26.579			10
ATOM		CB LEU	A 46	24.608			_
ATOM		CG LEU		23.642			
ATOM		CD1 LEU		24.089			
ATOM		CD2 LEU	A 46	22.275			
ATOM		M GLU	A 47	26.523			
ATOM		CA GLU .	A 47	26.910			
ATOM	297 (GLU :	A 47	28.140	-2.500		
ATOM		GLU 2	A 47	28.722	-3.203	28.500	
ATOM		B GLU	A 47	27.147	-4.206	26.204	
ATOM		G GLU A	A 47	27.386	-5.254	27.245	
ATOM		D GLU A	47	27.661	-6.560	26.524	
ATOM		E1 GLU A	47	26.741	-7.007		1.00 68.40
ATOM		E2 GLU F	47	28.856	-6.921	25.777	1.00 66.37
ATOM	304 N	SER A	48	28.992	-1.626	26.830 27.215	1.00 78.70
ATOM		A SER A	48	30.331	-1.518	27.789	1.00 27.50
ATOM	306 C	SER A	48	30.108	-0.555	28.926	1.00 25.23
ATOM	307 o	SER A	48	31.124	-0.058	29.462	1.00 26.91
ATOM	308 C			31.116	-0.990	26.621	1.00 33.39
ATOM	309 0		48	31.294	0.422	26.483	1.00 21.90
ATOM	310 N		49	28.826	-0.101	28.995	1.00 27.87
ATOM	311 C		49	28.542	0.955	29.956	1.00 25.04
ATOM	312 C	HIS A	49	27.480	0.461	30.950	1.00 19.72
ATOM	313 0	HIS A	49	27.186	1.089	31.898	1.00 22.55 1.00 27.93
ATOM	314 CE		49	28.094	2.197	29.463	-
ATOM	315 C		49	28.806	3.036	28.520	1.00 16.13 1.00 39.79
ATOM	316 NI		49	29.564	4.058	28.953	1.00 39.79 1.00 45.66
ATOM	317 CE		49	28.776	3.070	27.197	
ATOM	318 CE		49	30.028	4.750	27.979	
ATOM	319 NE		49	29.544	4.139	26.934	
ATOM	320 N	ILE A	50	27.009	-0.703	30.715	
ATOM	321 CA		50	25.874	-1.129	31.415	1.00 18.34
ATOM	322 C	ILE A	50	25.917	-2.629	31.146	1.00 19.89
ATOM	323 0		50	25.322	-3.023	30.168	1.00 26.29 1.00 25.33
ATOM	324 CB		50	24 505	-0.535	31.008	1.00 25.33 1.00 10.50
ATOM		1 ILE A	50	24.340		31.292	
ATOM		2 ILE A	50			31.697	1.00 4.97 1.00 12.96
ATOM	327 CD		50	23.413		30.602	1.00 12.96
ATOM	328 N	ARG A	51			32.066	1.00 16.65
ATOM	329 CA		51			32.107	1.00 31.77
ATOM	330 C	ARG A	51			32.170	
ATOM	331 0	ARG A	51			31.512	1.00 32.68 1.00 37.16
ATOM	332 N	ASN A	52			32.788	1.00 37.16
ATOM	333 CA	ASN A	52				1.00 28.48
ATOM ATOM	334 C	ASN A	52				1.00 26.39
AIOM	335 o	ASN A	52				1.00 27.75
						-3.000	1.00 20.08

ATOM	336	CB	ASN		52	22.750	-5.884	34.232	1.00	34.86
ATOM	337	CG	ASN	Α	52	21.637	-6.879	34.271	1.00	39.54
ATOM	338	OD1	ASN	Α	52	20.781	-6.541	35.095	1.00	54.31
ATOM	339	ND2	ASN		52	21.611	-7.954	33.503	1.00	48.82
ATOM	340	N	ILE	Α	53	22.127	-5.699	30.800	1.00	24.42
ATOM	341	CA	ILE	Α	53	21.261	-5.092	29.772		20.15
ATOM	342	С	ILE	Α	53	20.585	-6.151	28.912		
ATOM	343	0	ILE	Α	53	21.020	-7.349	28.917	1.00	18.01
ATOM	344	CB	ILE	Α	53	22.245	-4.297	28.880		14.09
ATOM	345	CG1			53	21.682	-3.257	27.936		
ATOM	346	CG2			53	22.907	-5.321	27.946	1.00	16.37
ATOM	347	CD1			53	22.877	-2.315	27.622		
ATOM	348	N	TRP		54	19.447	-5.880	28.383		15.19
ATOM	349	CA	TRP		54	18.804	-6.889	27.567		
ATOM	350	C	TRP		54	18.803	-6.230	26.151		19.82
ATOM	351	ō	TRP		54	18.340	-5.059	25.985		18.37
ATOM	352	CB	TRP		54	17.364	-7.046	27.998		23.18
ATOM	353	CG	TRP		54	16.949	-7.932	29.100		24.57
ATOM	354	CD1	TRP		54	17.757	-8.727			24.37
ATOM	355	CD2	TRP		54	15.595	-8.164	29.895		
ATOM	356	NE1	TRP		54	17.004		29.603	1.00	
ATOM	357	CE2	TRP		54		-9.372	30.858	1.00	
ATOM	358	CE3	TRP			15.692	-9.039	30.700	1.00	
		CZ2			54	14.358	-7.633	29.243	1.00	
ATOM	359		TRP		54	14.611	-9.442	31.432	1.00	
ATOM	360	CZ3	TRP		54	13.316	-8.042	30.009	1.00	
ATOM	361	CH2	TRP		54	13.451	-8.916	31.068	1.00	
ATOM	362	N	ILE		55	19.063	-7.152	25.204	1.00	
ATOM	363	CA		Α	55	19.178	-6.655	23.838	1.00	
ATOM	364	С	ILE		55	18.091	-7.215	22.962	1.00	
ATOM	365	0	ILE		55	17.955	-8.378	22.680	1.00	7.34
ATOM	366	CB		A	55	20.546	-6.962	23.201	1.00	16.44
ATOM	367	CG1	ILE		55	21.939	-6.409	23.702	1.00	8.75
ATOM	368	CG2	ILE		55	20.384	-6.460	21.750	1.00	21.77
ATOM	369	CD1	ILE	Α	55	21.767	-5.582	24.863	1.00	16.23
ATOM	370	N	GLN	Α	56	17.226	-6.412	22.390	1.00	9.67
ATOM	371	CA	GLN	Α	56	16.161	-7.016	21.619	1.00	10.90
ATOM	372	С	GLN	Α	56	16.432	-6.621	20.143	1.00	13.08
ATOM	373	0	GLN	Α	56	16.402	-5.393	19.953	1.00	
ATOM	374	CB	GLN	Α	56	14.786	-6.542	22.014	1.00	
ATOM	375	CG	GLN	Α	56	13.653	-7.256	21.316	1.00	
ATOM	376	CD	GLN		56	13.789	-8.741	21.351	1.00	
ATOM	377	OE1	GLN		56	13.610	-9.379	20.324	1.00	9.56
ATOM	378	NE2	GLN		56	14.119	-9.221	22.544	1.00	
ATOM	379	N	GLY		57	16.288	-7.645	19.216	1.00	6.84
ATOM	380	CA	GLY		57	16.174	-7.019	17.841	1.00	
ATOM	381	C	GLY		57	14.740	-7.085	17.267		13.72
ATOM	382	0	GLY		57	14.124	-8.016	17.752		12.70
ATOM	383	N	VAL		58	14.068	-6.264	16.525		
		- 1	*******	4 1	50	14.000	0.204	10.323	1.00	12.13

ATOM	384	CA	VAL		58		2.739		16.070		11.16
ATOM	385	С	VAL		58		2.715	-7.246	14.893		14.85
ATOM	386	0	VAL		58		3.234	-6.891	13.849		18.64
ATOM	387	CB	VAL		58		2.262	-4.984	15.352	1.00	6.54
ATOM	388	CG1			58		0.894	-4.974	14.731	1.00	5.89
ATOM	389	CG2	VAL	Α	58	1	2.650	-3.840	16.331	1.00	5.86
ATOM	390	N	GLY	Α	59	1	2.209	-8.465	15.008	1.00	21.96
ATOM	391	CA	GLY	Α	59	1	2.120	-9.385	13.874	1.00	17.81
ATOM	392	С	GLY	Α	59	1	0.645	-9.561	13.550	1.00	23.35
ATOM	393	0	GLY	A	59		9.919	-8.579	13.249	1.00	27.99
ATOM	394	N	GLY	Α	60	1	0.166	-10.805	13.623	1.00	18.75
ATOM	395	CA	GLY	A	60		8.841	-11.142	13.285	1.00	11.46
ATOM	396	С	GLY	Α	60		8.550	-10.833	11.851	1.00	14.56
ATOM	397	0	GLY	Α	60		9.160	-11.439	11.003	1.00	16.32
ATOM	398	N	PRO	A	61			-10.103	11.612	1.00	12.10
ATOM	399	CA	PRO	Α	61		7.123	-9.774	10.250	1.00	14.70
ATOM	400	С	PRO		61		8.230	-8.941	9.570		22.17
ATOM	401	0	PRO		61		8.143	-8.758	8.344	1.00	
ATOM	402	СВ	PRO		61		5.911	-8.860	10.332	1.00	
ATOM	403	CG	PRO		61		5.880	-8.514	11.784	1.00	
ATOM	404	CD	PRO		61		6.723	-9.417	12.576	1.00	
ATOM	405	N	TYR		62		9.162	-8.257	10.292	1.00	
ATOM	406	CA	TYR		62		9.973	-7.242	9.674	1.00	
ATOM	407	C	TYR		62		1.133	-7.907	9.047	1.00	
ATOM	408	0	TYR		62		2.132	-8.213	9.691	1.00	
ATOM	409	CB	TYR		62		0.504	-6.401	10.803	1.00	
ATOM	410	CG	TYR		62		1.461	-5.421	10.236	1.00	
ATOM	411	CD1	TYR		62		1.343	-4.920	9.032	1.00	
ATOM	412	CD2	TYR		62		2.465	-4.971	10.969	1.00	
ATOM	413	CE1	TYR		62		2.206	-3.997	8.506	1.00	19.28
ATOM	414	CE2	TYR		62		3.438	-4.101	10.490	1.00	
ATOM	415	CZ	TYR		62		3.327	-3.571	9.186	1.00	
ATOM	416	OH	TYR		62		4.320	-2.649	8.791	1.00	
ATOM	417	N	ASP		63		0.998	-8.419	7.816	1.00	
ATOM	418	CA	ASP		63		2.137	-9.011	7.081	1.00	
ATOM	419	CA	ASP		63		3.027	-7.973	6.453		17.97
					63		3.628				14.94
ATOM	420	O	ASP					-8.442	5.512		
ATOM	421	CB	ASP		63		1.474	-9.873	6.015		17.16
ATOM	422	CG	ASP		63		0.563				27.75
ATOM	423		ASP		63		0.049				34.11
ATOM	424		ASP		63		0.300				44.13
ATOM	425	N	ALA		64		3.089				15.36
ATOM	426	CA	ALA		64		4.054				17.14
ATOM	427	С	ALA		64		4.118				21.10
ATOM	428	0	ALA		64		5.193				23.12
ATOM	429	CB	ALA		64		5.458				20.45
ATOM	430	N	ALA		65		2.946	-6.009	4.006		22.21
ATOM	431	CA	ALA	A	65	1	2.817	-6.072	2.565	1.00	21.81

ATOM	432	С	ALA	Α	65	13.143	-4.857	1.745	1.00	21.76
ATOM	433	0	ALA	A	65	12.855	-3.801	2.229		23.60
ATOM	434	СВ	ALA	Α	65	11.384	-6.390	2.364	1.00	17.31
ATOM	435	N	LEU	A	66	13.401	-4.866	0.402	1.00	21.48
ATOM	436	CA	LEU	A	66	13.763	-3.581	-0.216	1.00	13.20
ATOM	437	С	LEU	Α	66	12.469	-2.913	-0.452	1.00	13.90
ATOM	438	0	LEU	A	66	12.548	-1.767	-0.197	1.00	11.85
ATOM	439	CB	LEU	Α	66	14.593	-3.602	-1.470	1.00	3.92
ATOM	440	CG	LEU	A	66	15.891	-4.308	-1.191	1.00	9.05
ATOM	441	CD1	LEU	Α	66	16.509	-4.725	-2.438	1.00	12.78
ATOM	442	CD2	LEU	Α	66	16.569	-3.119	-0.580	1.00	13.44
ATOM	443	N	ALA	A	67	11.413	-3.625	-0.801	1.00	14.94
ATOM	444	CA	ALA	A	67	10.253	-2.759	-1.277		12.42
ATOM	445	С	ALA	A	67	9.626	-1.879	-0.224		14.21
ATOM	446	0	ALA	A	67	9.218	-0.818	-0.643		14.29
ATOM	447	CB	ALA	Α	67	9.089	-3.588	-1.781	1.00	3.90
ATOM	448	N	THR	A	68	9.494	-2.409	1.006	1.00	12.11
ATOM	449	CA	THR		68	8.780	-1.647	1.997	1.00	11.77
ATOM	450	С	THR		68	9.242	-0.214	2.219		13.05
ATOM	451	0	THR		68	8.597	0.683	2.766		11.13
ATOM	452	CB	THR		68	8.892	-2.488	3.241		13.93
ATOM	453	OG1	THR		68	10.145	-3.150	3.224		27.44
ATOM	454	CG2	THR		68	7.783	-3.459	3.087		13.39
ATOM	455	N	ASN	A	69	10.450	-0.057	1.808	1.00	7.59
ATOM	456	CA	ASN	Α	69	11.020	1.236	1.791	1.00	8.76
ATOM	457	С	ASN	A	69	10.095	2.165	1.047	1.00	
ATOM	458	0	ASN	Α	69	9.950	3.345	1.305	1.00	5.30
ATOM	459	СВ	ASN	Α	69	12.461	1.251	1.231	1.00	5.54
ATOM	460	CG	ASN	Α	69	13.374	1.207	2.398	1.00	15.08
ATOM	461	OD1	ASN	A	69	13.307	2.124	3.275	1.00	31.90
ATOM	462	ND2	ASN	A	69	14.048	0.099	2.360	1.00	4.51
ATOM	463	N	PHE	Α	70	9.390	1.656	0.079	1.00	19.09
ATOM	464	CA	PHE	Α	70	8.552	2.619	-0.631		21.80
ATOM	465	С	PHE	Α	70	7.157	2.836	-0.123	1.00	23.36
ATOM	466	0	PHE	A	70	6.509	3.717	-0.724		25.74
ATOM	467	CB	PHE	Α	70	8.547	2.386	-2.082	1.00	17.38
ATOM	468	CG	PHE	Α	70	9.870	2.360	-2.770	1.00	15.72
ATOM	469	CD1	PHE	A	70	10.080	3.430	-3.576	1.00	5.15
ATOM	470	CD2	PHE	A	70	10.702	1.245	-2.497	1.00	7.61
ATOM	471	CE1	PHE	A	70	11.268	3.330	-4.191	1.00	16.05
ATOM	472	CE2	PHE	Α	70	11.913	1.267	-3.168		22.23
ATOM	473	CZ	PHE	Α	70	12.199	2.314	-4.016	1.00	9.57
ATOM	474	N	LEU	Α	71	6.765	2.246	1.034	1.00	25.53
ATOM	475	CA	LEU	A	71	5.506	2.725	1.599		24.24
MOTA	476	С	LEU	A	71	5.649	4.037	2.343	1.00	27.91
ATOM	477	0	LEU	A	71	6.694	4.521	2.750		28.86
ATOM	478	CB	LEU	Α	71	5.150	1.635	2.535		19.99
MOTA	479	CG	LEU	Α	71	5.003	0.342	1.873	1.00	16.09

ATOM	480	CD1	LEU	Α	71	4.	879	-0.	764	2	.885	1.00	18.12
ATOM	481	CD2	LEU	Α	71		786		546		.000		18.24
ATOM	482	N	PRO	Α	72		535		663		.529		33.01
ATOM	483	CA	PRO	Α	72		389		888		.311	1.00	
ATOM	484	С	PRO	Α	72		865		590		.778	1.00	
ATOM	485	0	PRO		72		619		512		.331	1.00	
ATOM	486	CB	PRO		72		983		453		.095	1.00	
ATOM	487	CG	PRO		72		224		189		.827	1.00	
ATOM	488	CD	PRO		72		188		093		.380	1.00	
ATOM	489	N	ARG		73		601		610		.221	1.00	
ATOM	490	CA	ARG		73		325		547		.408	1.00	
ATOM	491	С	ARG		73		613		755		.321		21.78
ATOM	492	0	ARG		73		360		950		.304		29.61
ATOM	493	CB	ARG		73		469		978		.549	1.00	
ATOM	494	CG	ARG		73		575		998		.155	1.00	
ATOM	495	CD	ARG		73		818		793		.360	1.00	
ATOM	496	NE	ARG		73		222		460		.392	1.00	
ATOM	497	CZ	ARG		73		391		312		.713		42.26
ATOM	498	NH1	ARG		73		145		288		.555		26.57
ATOM	499	NH2	ARG		73		320		144		.883	1.00	
ATOM	500	N	GLY		74		368		909		.326	1.00	8.42
ATOM	501	CA	GLY		74		L20		291		.332	1.00	5.06
ATOM	502	С	GLY		74	9.2			858		.508		12.74
ATOM	503	0	GLY		74	10.2			286		.317		16.46
ATOM	504	N	THR		75	8.1			321		906		12.82
ATOM	505	CA	THR		75	8.0			869		.008	1.00	11.14
ATOM	506	С	THR		75	6.6			428		134	1.00	10.64
ATOM	507	0	THR	Α	75	5.7			231		949	1.00	9.36
ATOM	508	СВ	THR		75	8.8			398		.219	1.00	6.97
ATOM	509	OG1	THR	Α	75	8.9		-0.			125	1.00	5.64
ATOM	51 0	CG2	THR	Α	75	8.1			865		603	1.00	6.30
ATOM	511	N	SER	A	76	6.4		-0.			259	1.00	10.07
MOTA	512	CA	SER	Α	76	5.0		-1.			354	1.00	13.33
ATOM	513	C	SER		76	4.4		-1.			747	1.00	21.87
ATOM	514	0	SER	Α	76	5.2		-1.			679	1.00	24.22
MOTA	515	CB	SER	Α	76	5.0		-2.			083	1.00	4.81
MOTA	516	OG	SER	Α	76	5.3		-3.			107		16.98
MOTA	517	N	GLN	Α	77	3.0		-1.			911		24.90
ATOM	518	CA	GLN		77	2.4		-1.			166		23.85
ATOM	519	С	GLN	A	77	2.6		-2.			015		19.58
ATOM	520	0	GLN	Α	77	2.9		-2.			203		15.15
ATOM	521	CB	GLN	Α	77	0.9		-0.			217		32.64
ATOM	522	CG	GLN		77	0.5		-0.			642		49.56
MOTA	523	CD	GLN	Α	77	0.6			785		194		65.91
ATOM	524	OE1	GLN	A	77	0.9			869		356		66.06
ATOM	525	NE2	GLN	A	77	0.4			750		350		68.91
ATOM	526	N	ALA	A	78	2.7		-3.			402		15.90
ATOM	527	CA	ALA	A	78	3.0		-4.			073		19.47

ATOM	528	С	ALA		78	4.381	-4.332	10.819		24.48
ATOM	529	0	ALA		78	4.389	-4.729	11.983		26.91
ATOM	530	CB	ALA		78	3.390	-5.808	9.336		17.23
ATOM	531	N	ASN		79	5.350	-3.863	10.093		21.58
ATOM	532	CA	ASN		79	6.602	-3.576	10.774	1.00	20.62
ATOM	533	С	ASN	Α	79	6.480	-2.673	11.969	1.00	20.93
ATOM	534	0	ASN	A	79	6.975	-2.944	13.053	1.00	15.52
ATOM	535	CB	ASN	Α	79	7.474	-3.069	9.670	1.00	24.79
ATOM	536	CG	ASN	A	79	7.933	-4.238	8.824	1.00	28.76
ATOM	537	OD1	ASN	Α	79	7.867	-5.439	9.091	1.00	25.30
ATOM	538	ND2	ASN	Α	79	8.488	-3.891	7.660	1.00	24.90
ATOM	539	N	ILE	A	80	5.731	-1.611	11.936	1.00	15.93
ATOM	540	CA	ILE	A	80	5.586	-0.574	12.924	1.00	17.00
ATOM	541	С	ILE	Α	80	4.925	-1.187	14.118		
ATOM	542	0	ILE	A	80	5.234	-0.939	15.264		18.79
ATOM	543	CB	ILE	A	80	4.756	0.629	12.436		11.98
ATOM	544	CG1	ILE		80	5.627	1.124	11.297	1.00	9.50
ATOM	545	CG2	ILE		80	4.379	1.728	13.354		16.27
ATOM	546	CD1	ILE		80	5.007	2.071	10.424	1.00	8.15
ATOM	547	N	ASP		81	4.017	-2.019	13.708		
ATOM	548	CA	ASP		81	3.304	-2.778	14.728		15.15
ATOM	549	C	ASP		81	4.147	-3.711	15.510		15.77
ATOM	550	Õ	ASP		81	4.084	-3.697	16.695		15.82
ATOM	551	CB	ASP		81	2.291	-3.438	13.868		26.36
ATOM	552	CG	ASP		81	1.065	-2.530	13.790	1.00	
ATOM	553	OD1			81	1.105	-1.355	14.226	1.00	
ATOM	554		ASP		81	0.061	-3.125	13.222	1.00	
ATOM	555	N	GLU		82	5.148	-4.447	15.096		16.07
ATOM	556	CA	GLU		82	5.984	-5.318	15.882		14.77
ATOM	557	C	GLU		82	6.839	-4.355	16.667		19.33
ATOM	558	Ö	GLU		82	7.315	-4.708	17.752	1.00	
ATOM	559	СВ	GLU		82	6.998	-6.031	15.064		13.20
ATOM	560	CG	GLU		82	7.792	-7.239	15.476	1.00	
ATOM	561	CD	GLU		82	6.767	-8.114	16.185		29.68
ATOM	562	OE1	GLU		82	5.666	-7.670	16.403		26.63
ATOM	563	OE2			82	7.273	-9.181	16.411		33.08
ATOM	564	N	GLY		83	7.228	-3.227	16.199	1.00	
ATOM	565	CA	GLY		83	8.033	-2.428	17.140	1.00	
ATOM	566	C	GLY		83	7.238	-2.018	18.366	1.00	
ATOM	567	0	GLY		83	7.561	-2.103	19.528	1.00	
ATOM	568	N	LYS		84	6.093	-1.408	18.114		18.72
ATOM	569	CA	LYS		84					
ATOM	570	CA		A	84	5.050	-1.146	19.096		16.90
ATOM	571		LYS		84	4.893	-2.337	20.057		17.74
ATOM	572	O CB	LYS		84	4.962	-2.265	21.295		14.31
ATOM	573	CB				3.799	-0.872	18.307		14.62
ATOM	574	CG		A n	84	3.535	0.565	18.291		19.30
		CD	LYS		84	2.787	1.013	17.044		34.24
ATOM	575	CE	LYS	А	84	1.568	1.902	17.337	1.00	31.10

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ATOM	576	NZ	LYS		84	0.346	1.226	16.827		48.42
ATOM	577	N	ARG		85	4.617	-3.506	19.519		18.50
ATOM	578	CA	ARG		85	4.583	-4.705	20.280		19.04
ATOM	579	С	ARG		85	5.677	-4.733	21.308		19.63
ATOM	580	0 .	ARG	Α	85	5.442	-5.192	22.383	1.00	19.24
ATOM	581	CB	ARG	Α	85	4.740	-5.979	19.464	1.00	14.74
ATOM	582	CG	ARG	A	85	3.843	-7.094	19.887	1.00	8.85
ATOM	583	CD	ARG	A	85	4.146	-8.554	19.705	1.00	7.20
ATOM	584	NE	ARG	Α	85	5.483	-8.898	19.194	1.00	20.30
ATOM	585	CZ	ARG		85	6.170	-9.705	19.899	1.00	18.19
ATOM	586	NH1	ARG		85	5.627	-10.161	21.040		34.03
ATOM	587	NH2	ARG		85	7.345	-9.979	19.555	1.00	
ATOM	588	N	LEU		86	6.901	-4.586	20.956	1.00	
ATOM	589	CA	LEU		86	8.006	-4.792	21.873	1.00	
ATOM	590	C	LEU		86	8.044	-3.637	22.803		20.73
ATOM	591	0	LEU		86	8.155	-3.037	23.925	1.00	22.18
	592	CB			86		-4.932			
ATOM			LEU			9.333		21.168	1.00	6.67
ATOM	593	CG	LEU		86	9.358	-6.241	20.282	1.00	
ATOM	594	CD1	LEU		86	10.546	-6.054	19.287	1.00	
ATOM	595	CD2	LEU		86	9.362	-7.516	21.020	1.00	5.17
ATOM	596	N		A	87	7.700	-2.446	22.529	1.00	
ATOM	597	CA	PHE	A	87	7.850	-1.416	23.492	1.00	
MOTA	598	С	PHE	A	87	6.939	-1.805	24.618	1.00	
ATOM	599	0	PHE	A	87	7.082	-1.565	25.839		30.36
ATOM	600	CB	PHE	Α	87	7.498	-0.118	22.846		15.81
ATOM	601	CG	PHE	Α	87	8.661	0.503	22.128		22.72
ATOM	602	CD1		Α	87	9.625	1.163	22.795	1.00	
ATOM	603	CD2		A	87	8.800	0.446	20.774	1.00	24.19
ATOM	604	CE1	PHE	Α	87	10.699	1.781	22.220	1.00	26.46
MOTA	605	CE2	PHE	A	87	9.871	0.991	20.153	1.00	29.24
ATOM	606	CZ	PHE	Α	87	10.827	1.669	20.849	1.00	20.81
ATOM	607	N	ALA	Α	88	5.862	-2.422	24.266	1.00	29.15
ATOM	608	CA	ALA	Α	88	4.772	-2.699	25.195	1.00	22.92
ATOM	609	С	ALA	Α	88	5.186	-3.837	26.068	1.00	22.03
ATOM	610	0	ALA	A	88	4.974	-3.879	27.284	1.00	
ATOM	611	CB	ALA		88	3.551	-2.803	24.299		22.13
ATOM	612	N	LEU	Α	89	5,649	-4.897	25.531		19.16
ATOM	613	CA	LEU		89	6.188	-6.032	26.208	1.00	19.29
ATOM	614	С	LEU		89	7.250		27.133		22.06
ATOM	615	Ō	LEU		89	7.449	-6.050	28.177		20.49
ATOM	616	CB	LEU		89	7.021	-6.863	25.221		18.41
ATOM	617	CG	LEU		89	7.477	-8.167	25.834		20.45
ATOM	618		LEU		89	6.326	-8.707	26.627		17.22
ATOM	619		LEU		89	8.060	-9.057	24.769		18.83
ATOM	620	N	ALA		90	8.124				
ATOM		CA						26.722		22.80
	621 622		ALA		90	9.027	-4.137 -3.499	27.701		24.14
ATOM	622	С	ALA		90	8.237		28.849		23.63
ATOM	623	0	ALA	A	90	8.414	-3.835	30.071	1.00	22.73

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ATOM	624	CB	ALA		90	10.080	-3.253	27.139	1.00 7.74
ATOM	625	N	ASN		91	7.457	-2.445	28.732	1.00 25.45
ATOM	626	CA	ASN	Α	91	6.665	-1.979	29.870	1.00 27.25
ATOM	627	С	ASN	Α	91	5.847	-2.996	30.656	1.00 30.97
ATOM	628	0	ASN	Α	91	5.346	-2.884	31.768	1.00 27.64
ATOM	629	CB	ASN	Α	91	5.560	-1.206	29.125	1.00 29.14
ATOM	630	CG	ASN	Α	91	4.946	-0.345	30.216	1.00 31.73
ATOM	631		ASN		91	3.845	-0.692	30.645	1.00 46.76
ATOM	632		ASN		91	5.641	0.629	30.643	1.00 29.03
ATOM	633	N	GLN		92	5.369	-4.008	29.969	1.00 35.37
ATOM	634	CA	GLN		92	4.702	-5.141	30.591	1.00 35.55
ATOM	635	C	GLN		92	5.619	-6.072	31.352	1.00 34.28
	636	0	GLN		92	5.227	-6.519	32.440	1.00 34.28
ATOM									
MOTA	637	CB	GLN		92	3.866	-5.903	29.573	1.00 54.94
MOTA	638	CG	GLN		92	2.689	-6.698	30.142	1.00 78.63
ATOM	639	CD	GLN		92	2.806	-8.167	29.805	1.00 93.87
ATOM	640	OE1	GLN		92	3.597	-8.840	30.475	1.00 96.99
MOTA	641	NE2	GLN		92	2.083	-8.696	28.824	1.00 97.81
MOTA	642	N	LYS	Α	93	6.859	-6.403	31.050	1.00 31.97
MOTA	643	CA	LYS	A	93	7.675	-7.204	31.972	1.00 25.22
MOTA	644	С	LYS	Α	93	8.381	-6.298	33.015	1.00 24.68
ATOM	645	0	LYS	Α	93	8.716	-6.793	34.075	1.00 32.13
ATOM	646	СВ	LYS	Α	93	8.673	-7.980	31.148	1.00 10.86
ATOM	647	CG	LYS	Α	93	8.225	-8.963	30.159	1.00 24.26
ATOM	648	CD	LYS		93	9.362	-9.966	29.986	1.00 21.96
ATOM	649	CE	LYS		93		-10.718	28.658	1.00 23.78
ATOM	650	NZ	LYS		93		-11.805	28.300	1.00 25.87
ATOM	651	N	CYS		94	8.752	-5.096	32.774	1.00 16.62
ATOM	652	CA	CYS		94	9.752	-4.412	33.480	1.00 18.95
ATOM	653	C	CYS		94	9.512	-2.936	33.537	1.00 24.83
		0			94				
ATOM	654		CYS			10.184	-2.017	33.150	1.00 26.80
MOTA	655	CB	CYS		94	11.147	-4.691	32.911	1.00 3.14
ATOM	656	SG	CYS		94	11.618	-6.437	32.882	1.00 25.28
ATOM	657	N	PRO		95	8.403	-2.561	34.086	1.00 26.08
ATOM	658	CA	PRO		95	7.891	-1.202	33.878	1.00 26.11
ATOM	659	С	PRO		95	8.960	-0.259	34.299	1.00 27.32
MOTA	660	0	PRO		95	8.776	0.966	34.108	1.00 29.08
MOTA	661	CB	PRO	Α	95	6.609	-1.090	34.747	1.00 20.75
MOTA	662	CG	PRO	Α	95	6.587	-2.421	35.322	1.00 19.04
MOTA	663	CD	PRO	Α	95	7.363	-3.461	34.509	1.00 22.55
MOTA	664	N	ASN	Α	96	9.836	-0.776	35.193	1.00 31.44
ATOM	665	CA	ASN	Α	96	10.559	0.274	35.966	1.00 35.38
ATOM	666	С	ASN	Α	96	11.891	0.476	35.353	1.00 33.83
ATOM	667	0	ASN		96	12.599	1.359	35.684	1.00 33.31
ATOM	668	СВ	ASN		96	10.558	-0.099	37.429	1.00 53.70
MOTA	669	CG	ASN		96	9.238	0.342	38.026	1.00 61.69
ATOM	670	OD1			96	8.758	1.432	37.706	1.00 64.33
ATOM	671		ASN		96	8.676	-0.526	38.861	1.00 67.25
414 Of 1	O , 1	11112	TOM	41	20	0.070	0.520	20.001	1.00 07.23

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ATOM	672	N	THR		97	12.287	-0.409	34.507		30.32
ATOM	673	CA	THR		97	13.519	-0.367	33.794		22.83
ATOM	674	C	THR		97	13.404	0.493	32.534		22.44
ATOM	675	0	THR		97	12.446	0.779	31.816		21.14
ATOM	676	CB	THR		97	13.835	-1.851	33.705		25.87
ATOM	677	OG1			97	14.602	-1.915	32.528	1.00	38.91
ATOM	678	CG2			97	12.769	-2.901	33.621	1.00	24.22
ATOM	679	N	PRO		98	14.393	1.415	32.408	1.00	20.59
ATOM	680	CA	PRO	A	98	14.513	2.292	31.254	1.00	18.15
ATOM	681	С	PRO	A	98	14.882	1.494	29.978	1.00	16.07
ATOM	682	0	PRO	Α	98	15.622	0.462	29.934	1.00	17.19
ATOM	683	CB	PRO	Α	98	15.563	3.339	31.676	1.00	
ATOM	684	CG	PRO	Α	98	16.270	2.646	32.699		12.29
ATOM	685	CD	PRO	Α	98	15.735	1.331	33.046		12.02
ATOM	686	N	VAL	Α	99	14.322	2.107	28.940		13.81
ATOM	687	CA	VAL		99	14.225	1.544	27.632		14.02
ATOM	688	С	VAL		99	14.956	2.407	26.663		10.66
ATOM	689	Ō	VAL		99	14.716	3.679	26.712	1.00	6.90
ATOM	690	CB	VAL		99	12.673	1.343	27.335	1.00	2.87
ATOM	691	CG1			99	12.666	1.272	25.872		17.40
ATOM	692				99	12.442	-0.111	27.744	1.00	
ATOM	693	N	VAL			15.885	1.776	25.861		5.75
ATOM	694	CA	VAL			16.525			1.00	6.45
ATOM	695	C	VAL				2.755	24.900	1.00	9.61
ATOM	696	0	VAL			16.389	2.159	23.561		10.79
ATOM	697	CB	VAL			16.256	0.973	23.477	1.00	9.11
ATOM	698		VAL			17.877	3.260	25.197	1.00	8.05
ATOM						17.824	4.252	26.336	1.00	6.05
	699		VAL			18.853	2.053	25.591	1.00	6.68
ATOM	700	N	ALA			16.277	2.928	22.511		13.14
ATOM	701	CA	ALA			16.127	2.266	21.183	1.00	15.67
ATOM	702	C	ALA .			17.065	2.747	20.053	1.00	12.08
ATOM	703	0	ALA .			17.261	4.042	19.907	1.00	11.16
ATOM	704	CB	ALA .			14.685	2.609	20.812	1.00	6.57
ATOM	705	N	GLY .			17.218	1.787	19.099	1.00	7.53
ATOM	706	CA	GLY .			17.949	2.415	17.939	1.00	7.10
ATOM	707	С	GLY .			17.477	1.803	16.744	1.00	7.27
ATOM	708	0	GLY .	A	102	17.102	0.621	16.878	1.00	10.83
ATOM	709	N	GLY .	A	103	17.706	2.407	15.648	1.00	7.80
ATOM	710	CA	GLY .	A	103	17.446	1.745	14.356	1.00	5.33
MOTA	711	С	GLY :	A	103	18.303	2.211	13.180	1.00	7.56
ATOM	712	0	GLY .	A	103	18.785	3.340	13.227	1.00	6.88
ATOM	713	N	TYR .	A	104	18.490	1.387	12.139	1.00	7.09
ATOM	714	ÇA	TYR .			19.392	1.682	11.069	1.00	5.99
ATOM	715	С	TYR .			18.705	1.614	9.705	1.00	9.47
ATOM	716	0	TYR :			18.115	0.638	9.441	1.00	6.46
ATOM	717	CB	TYR :			20.592	0.797	11.079	1.00	5.40
ATOM	718	CG	TYR I			21.436	1.078	9.876	1.00	8.05
ATOM	719	CD1	TYR			21.708	2.302	9.352	1.00	5.91
		_			- · -	,00	2.002	2.372	1.00	J. JI

ATOM	720	CD2	TYR A		21.961	-0.044	9.172		.85
ATOM	721	CE1	TYR A		22.447	2.513	8.186	1.00 5	.61
ATOM	722	CE2	TYR A		22.751	0.052	8.072	1.00 7	.49
ATOM	723	CZ		104	22.972	1.377	7.608		.08
ATOM	724	OH	TYR A		23.795	1.509	6.479	1.00 14	.32
ATOM	725	N	SER A	105	18.939	2.975	8.852	1.00 18	.39
ATOM	726	CA	SER A	105	18.190	2.854	7.601	1.00 9	.66
ATOM	727	С	SER A	105	16.763	2.370	7.722	1.00 6	.10
ATOM	728	0	SER A	105	16.090	3.304	8.077	1.00 5	. 63
ATOM	729	CB	SER A	105	19.124	2.159	6.607	1.00 8	. 55
ATOM	730	OG	SER A	105	18.553	1.685	5.463	1.00 24	.30
ATOM	731	N	GLN A	106	16.241	1.405	7.079	1.00 9	. 93
ATOM	732	CA	GLN A	106	14.759	1.316	7.002	1.00 8	.25
ATOM	733	С	GLN A	106	14.453	1.089	8.473	1.00 8	.51
ATOM	734	0	GLN A	106	13.470	1.683	8.862	1.00 6	.31
ATOM	735	CB	GLN A	106	14.239	0.393	5.940	1.00 7.	. 45
ATOM	736	CG	GLN A	106	13.184	-0.528	6.465	1.00 18.	.04
ATOM	737	CD	GLN A	106	12.228	-1.220	5.581	1.00 16	. 87
ATOM	738	OE1	GLN A	106	11.024	-1.180	5.492	1.00 17.	. 59
ATOM	739	NE2	GLN A	106	12.643	-2.032	4.713		. 32
ATOM	740	N	GLY A		15.269	0.310	9.172		.13
ATOM	741	CA	GLY A		15.190	0.159	10.606		. 61
ATOM	742	С	GLY A		15.048	1.472	11.356		.27
ATOM	743	0	GLY F		14.219	1.511	12.290		. 52
ATOM	744	N	ALA A		15.653	2.637	11.033		. 44
ATOM	745	CA	ALA A		15.266	3.864	11.641		. 41
ATOM	746	С	ALA A		13.813	4.346	11.471	1.00 11.	
ATOM	747	0	ALA A		13.150	4.914	12.298	1.00 12.	
ATOM	748	CB	ALA A		16.121	5.006	11.170	1.00 13.	
ATOM	749	N	ALA A		13.321	4.312	10.267		.78
ATOM	750	CA	ALA A		12.056	4.685	9.861	1.00 10.	
ATOM	751	С	ALA A		11.093	3.858	10.727	1.00 12.	
ATOM	752	Ō	ALA A		10.016	4.391	11.035	1.00 14.	
ATOM	753	CB	ALA A		12.035	4.173	8.456	1.00 10.	
ATOM	754	N	LEU F		11.259	2.690	11.077		.34
ATOM	755	CA	LEU F		10.458	1.760	11.783	1.00 11.	
ATOM	756	C	LEU F		10.305	2.253	13.203	1.00 15.	
ATOM	757	Ō	LEU F		9.298	2.672	13.685	1.00 18	
ATOM	758	СВ	LEU F		11.031	0.319	11.634		. 52
ATOM	759	CG	LEU A		10.247	-0.801	12.258		.41
ATOM	760	CD1	LEU F		10.685	-2.233	11.862		.17
ATOM	761	CD2	LEU P		10.278	-0.659	13.783		. 25
ATOM	762	N	ILE A		11.397	2.373	13.907	1.00 15.	
ATOM	763	CA	ILE A		11.510	2.860	15.246	1.00 13.	
ATOM	764	C	ILE A		11.027	4.255	15.234		.39
ATOM	765	Ö	ILE A		10.404	4.636	16.241	1.00 12.	
ATOM	766	СВ	ILE F		12.977	2.814	15.685	1.00 12.	
ATOM	767	CG1	ILE F		13.222	1.279	15.805	1.00 13.	
111 011	, , ,	CGT	ع ندید		13.222	1.213	10.000	T.00 T4	. I J

ATOM	768	CG2			111	13.195	3.465	17.005	1.00 4.64
MOTA	769	CD1			111	12.410	0.887	17.002	1.00 14.88
ATOM	770	N	ALA	Α	112	11.309	5.170	14.341	1.00 11.00
ATOM	771	CA	ALA	Α	112	10.792	6.528	14.427	1.00 12.45
ATOM	772	С	ALA	Α	112	9.266	6.455	14.308	1.00 15.59
ATOM	773	0	ALA	Α	112	8.728	7.131	15.154	1.00 18.13
ATOM	774	CB	ALA	Α	112	11.334	7.505	13.486	1.00 5.70
ATOM	775	N	ALA	Α	113	8.575	5.572	13.587	1.00 12.85
ATOM	776	CA	ALA	Α	113	7.167	5.512	13.557	1.00 15.39
ATOM	777	С	ALA	Α	113	6.475	5.093	14.861	1.00 18.21
ATOM	778	0			113	5.498	5.750	15.226	1.00 14.59
ATOM	779	CB			113	6.678	4.562	12.500	1.00 17.63
ATOM	780	N			114	6.937	3.948	15.303	1.00 16.02
ATOM	781	CA.	ALA			6.483	3.218	16.412	1.00 16.43
ATOM	782	С	ALA			6.578	4.114	17.643	1.00 22.20
ATOM	783	Ō	ALA			5.673	4.321	18.426	1.00 18.94
ATOM	784	СВ	ALA			7.474	2.084	16.565	1.00 4.69
ATOM	785	N	VAL			7.722	4.836	17.744	1.00 22.46
ATOM	786	CA	VAL			7.855	5.499	19.064	1.00 20.88
ATOM	787	C	VAL			6.670	6.469	19.007	1.00 22.71
ATOM	788	O.	VAL			6.136	6.761	20.057	1.00 22.71
ATOM	789	СВ	VAL			9.279	6.090	19.137	1.00 19.61
ATOM	790	CG1	VAL			9.396	7.259	20.122	1.00 8.35
ATOM	791		VAL			10.245	5.016	19.562	1.00 13.91
ATOM	792	N	SER			6.467	7.085	17.828	1.00 23.59
ATOM	793	CA	SER			5.539	8.172	17.736	1.00 23.68
ATOM	794	C	SER			4.169	7.647	18.120	1.00 23.00
ATOM	795	0	SER			3.333	8.523	18.399	1.00 23.77
ATOM	796	СВ	SER			5.522	8.865	16.376	1.00 27.33
ATOM	797	OG	SER			5.168	8.043	15.277	
ATOM	798	N	GLU			3.859	6.397		
ATOM	799	CA	GLU			2.491		18.004	1.00 18.83
ATOM	800	CA	GLU				6.020	18.238	1.00 22.21
ATOM	801	0				2.461	5.474	19.653	1.00 30.46
ATOM	802		GLU			1.487	4.773	19.863	1.00 35.72
ATOM	803	CB CG	GLU GLU			1.977	4.902	17.343	1.00 21.63
						2.167	5.219	15.897	1.00 26.41
ATOM	804	CD	GLU			1.560	4.424	14.814	1.00 34.01
ATOM	805		GLU			0.912	3.440	15.046	1.00 32.59
ATOM	806		GLU			1.750	4.833	13.659	1.00 44.62
ATOM	807	N	LEU			3.438	5.570	20.512	1.00 34.45
ATOM	808	CA	LEU			3.326	5.006	21.812	1.00 33.64
ATOM	809	С	LEU			2.681	6.110	22.633	1.00 41.75
ATOM	810	0	LEU			2.594	7.267	22.370	1.00 39.90
ATOM	811	CB	LEU			4.600	4.668	22.392	1.00 29.44
ATOM	812	CG	LEU			5.628	3.891	21.645	1.00 26.36
ATOM	813		LEU			6.921	3.840	22.379	1.00 27.53
ATOM	814		LEU			5.110	2.520	21.536	1.00 20.69
MOTA	815	N	SER	A	119	2.076	5.794	23.726	1.00 48.86

ATOM	816	CA	SER			0.910	5.647	24.476		52.44
ATOM	817	С	SER	A	119	1.212	6.063	25.866		52.57
ATOM	818	0	SER	A	119	1.485	5.258	26.735	1.00	55.54
ATOM	819	CB	SER	Α	119	0.550	4.132	24.488		70.55
ATOM	820	OG	SER	Α	119	1.393	3.091	23.908	1.00	66.80
ATOM	821	N	GLY	Α	120	1.532	7.307	26.024	1.00	52.95
ATOM	822	CA	GLY	Α	120	1.910	7.761	27.382	1.00	53.35
ATOM	823	С	GLY	Α	120	2.944	7.109	28.291	1.00	49.09
ATOM	824	0			120	4.086	7.617	28.358	1.00	49.66
ATOM	825	N	ALA			2.526	6.129	29.102	1.00	42.97
ATOM	826	CA	ALA			3.477	5.574	30.022	1.00	40.72
ATOM	827	C	ALA			4.587	4.772	29.326	1.00	44.20
ATOM	828	Ō	ALA			5.749	4.803	29.711		45.42
ATOM	829	CB	ALA			2.965	4.542	30.903		36.34
ATOM	830	N	VAL			4.122	4.035	28.312		41.15
ATOM	831	CA	VAL			5.090	3.269	27.548		33.41
ATOM	832	C	VAL			5.870	4.168	26.652		28.48
	833	0	VAL			7.084	4.100	26.872		27.69
ATOM	834	CB	VAL			4.424	2.056	26.952		30.22
ATOM							1.997	27.098		28.03
ATOM	835	CG1 CG2				2.924		25.551		23.22
ATOM	836					4.891	1.836	26.177		23.16
ATOM	837	N	LYS			5.424	5.310			23.10
ATOM	838	CA	LYS			6.354	6.314	25.661		
ATOM	839	С	LYS			7.403	6.783	26.661		25.28
ATOM	840	0	LYS			8.524	7.224	26.449		29.01
ATOM	841	CB	LYS			5.561	7.502	25.100		23.54
ATOM	842	CG			123	6.171	8.573	24.277		26.71
ATOM	843	CD			123	5.400	9.775	23.888		43.07
ATOM	844	CE	LYS			4.953	9.783	22.461		59.59
ATOM	845	NZ	LYS			3.518	9.637	22.099		67.50
ATOM	846	N	GLU	A	124	6.977	6.991	27.918		27.95
ATOM	847	CA	GLU	Α	124	7.845	7.700	28.863		27.29
ATOM	848	С	GLU	Α	124	8.910	6.706	29.243		25.21
ATOM	849	0	GLU	Α	124	9.993	7.165	29.769	1.00	21.21
MOTA	850	CB	GLU	A	124	6.986	8.351	29.927	1.00	40.13
MOTA	851	CG	GLU	Α	124	7.588	8.609	31.295	1.00	57.40
ATOM	852	CD	GLU	Α	124	8.530	9.814	31.247	1.00	66.99
ATOM	853	OE1	GLU	Α	124	9.619	9.751	31.902	1.00	70.44
ATOM	854	OE2	GLU			7.949	10.652	30.502	1.00	73.84
ATOM	855	N			125	8.656	5.393	29.058	1.00	19.93
ATOM	856	CA	GLN			9.761	4.509	29.546	1.00	17.98
ATOM	857	С	GLN			10.865	4.556	28.521		24.28
ATOM	858	Ö	GLN			11.964	4.107	28.815		21.47
ATOM	859	СВ	GLN			9.225	3.178	29.844	1.00	9.13
ATOM	860	CG			125	9.901	2.001	30.299	1.00	9.05
ATOM	861	CD	GLN			9.211	0.719	30.129		19.33
ATOM	862	OE1			125	8.190	0.703	29.466		28.52
	863	NE2			125	9.662	-0.396	30.684		13.34
ATOM	003	NEZ	GTIN	М	120	9.002	-0.330	50.004	1.00	13.54

ATOM	864	N			126	10.593	5.188	27.319	1.00	25.30
ATOM	865	CA	VAL	Α	126	11.738	5.124	26.361	1.00	22.55
ATOM	866	С	VAL	Α	126	12.546	6.334	26.614	1.00	17.55
ATOM	867	0	VAL	A	126	12.109	7.408	26.329	1.00	12.79
ATOM	868	CB	VAL	Α	126	11.227	4.560	25.022	1.00	23.76
ATOM	869	CG1	VAL	Α	126	9.706	4.686	24.946	1.00	23.77
ATOM	870	CG2	VAL	Α	126	11.795	5.081	23.743	1.00	23.81
ATOM	871	N	LYS	Α	127	13.726	6.233	27.264	1.00	16.41
ATOM	872	CA	LYS	Α	127	14.462	7.494	27.639	1.00	18.18
ATOM	873	С	LYS	Α	127	15.239	8.063	26.488	1.00	
ATOM	874	0	LYS	A	127	15.812	9.103	26.680	1.00	
ATOM	875	CB	LYS	Α	127	15.401	7.148	28.792	1.00	
ATOM	876	CG	LYS	Α	127	14.770	6.110	29.713	1.00	
ATOM	877	CD			127	13.435	6.726	30.064		33.86
ATOM	878	CE			127	12.779	6.612	31.399	1.00	
ATOM	879	NZ			127	12.279	7.863	31.993	1.00	
ATOM	880	N			128	15.522	7.281	25.416		20.56
ATOM	881	CA			128	16.280	7.948	24.306		20.72
ATOM	882	C	GLY		*	16.358	7.104	23.063	1.00	17.71
ATOM	883	0	GLY			16.168	5.901	23.226	1.00	16.66
ATOM	884	N	VAL			16.451	7.725	21.892	1.00	
ATOM	885	CA	VAL			16.497	6.872	20.691		13.82
ATOM	886	C	VAL			17.519	7.371	19.719	1.00	8.35
ATOM	887	0	VAL			17.602	8.553	19.719	1.00	3.85
ATOM	888	CB	VAL			15.192	6.426	20.054	1.00	
ATOM	889	CG1	VAL			14.007	7.041			
ATOM	890	CG2	VAL					20.726	1.00	6.50
ATOM	891	N N	ALA			15.051	6.729	18.571	1.00	10.03
						18.455	6.398	19.363	1.00	8.05
ATOM	892	CA	ALA			19.430	6.845	18.344	1.00	7.55
ATOM	893	С	ALA			19.078	6.293	16.958	1.00	11.17
ATOM	894	0	ALA			18.755	5.145	16.849	1.00	15.74
ATOM	895	CB	ALA			20.781	6.391	18.603	1.00	5.89
ATOM	896	N	LEU			18.911	6.953	15.892	1.00	7.36
ATOM	897	CA	LEU			18.635	6.625	14.553	1.00	7.70
ATOM	898	C	LEU			19.876	6.908	13.661	1.00	12.02
ATOM	899	0	LEU			20.436	8.033	13.604	1.00	6.80
ATOM	900	CB	LEU			17.604	7.713	14.102	1.00	8.40
ATOM	901	CG	LEU			16.160	7.830	14.575	1.00	6.67
ATOM	902	CD1	LEU			15.391	8.957	13.981	1.00	4.49
ATOM	903	CD2	LEU			15.481	6.488	14.324	1.00	5.12
ATOM	904	N	PHE			20.271	6.009	12.802	1.00	11.56
ATOM	905	CA	PHE	Α	132	21.422	6.183	11.908	1.00	10.44
ATOM	906	С	PHE			20.965	6.013	10.478	1.00	8.46
ATOM	907	0	PHE			20.175	5.101	10.097	1.00	11.04
MOTA	908	CB	PHE			22.217	4.931	12.282	1.00	10.56
ATOM	909	CG	PHE	Α	132	22.693	4.830	13.714		16.38
ATOM	910	CD1	PHE	Α	132	21.951	4.029	14.542		13.36
ATOM	911	CD2	PHE	A	132	23.860	5.489	14.213	1.00	15.12

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ATOM	912		PHE A			22.342	3.911	15.889	1.00	14.91
ATOM	913		PHE A			24.176	5.323	15.513	1.00	18.02
ATOM	914	CZ	PHE A	132		23.426	4.530	16.403	1.00	15.09
ATOM	915	N	GLY A			21.431	6.876	9.580	1.00	7.35
ATOM	916	CA	GLY A	133		21.026	6.893	8.148	1.00	5.86
ATOM	917	С	GLY A	133		19.503	6.919	8.061	1.00	
ATOM	918	0	GLY A	133		18.890	5.926	7.593	1.00	
ATOM	919	N	TYR A	134		18.926	8.070	8.532	1.00	
ATOM	920	CA	TYR A	134		17.455	8.022		1.00	7.40
ATOM	921	С	TYR A	134		16.647	8.365	7.584	1.00	
ATOM	922	0	TYR A	134	•	16.785	9.513		1.00	5.85
ATOM	923	CB	TYR A			17.161	9.128	9.836	1.00	7.27
ATOM	924	CG	TYR F			15.842	9.393	10.391	1.00	
ATOM	925	CD1	TYR A			14.889	8.437	10.312	1.00	6.65
ATOM	926	CD2	TYR A			15.661	10.651	10.948	1.00	
ATOM	927	CE1	TYR A			13.657	8.690	10.821	1.00	9.05
ATOM	928	CE2	TYR A			14.408	10.928	11.467		12.89
ATOM	929	CZ	TYR A			13.428	9.923	11.423		14.22
ATOM	930	OH	TYR A			12.146	10.110	11.975		12.41
ATOM	931	N	THR A			15.811	7.398	7.139		11.51
ATOM	932	CA	THR A			15.229	7.581	5.789		7.71
ATOM	933	С	THR A			14.082	8.530	5.825		10.36
ATOM	934	O	THR A			13.845	8.878	4.727		11.26
ATOM	935	СВ	THR A			14.772	6.394	4.967		12.02
ATOM	936	OG1	THR A			13.821	5.399	5.398		22.81
ATOM	937	CG2	THR A			15.828	5.332	4.712		14.88
ATOM	938	N	GLN A			13.632	9.105	6.928		15.28
ATOM	939	CA	GLN A			12.596	10.134	6.968		16.48
ATOM	940	C	GLN A			13.102	11.418	7.646		17.46
ATOM	941	ō	GLN A			12.292	12.231	8.035		12.82
MOTA	942	СВ	GLN A			11.336	9.671	7.701		5.71
ATOM	943	CG	GLN A			11.178	8.191	7.263		13.60
ATOM	944	CD	GLN A			10.504	8.264	5.932		14.65
MOTA	945	OE1				9.587	9.102	5.986		23.99
ATOM	946	NE2	GLN A			10.852	7.529	4.914		14.68
ATOM	947	N	ASN A			14.421	11.532	7.566		18.52
ATOM	948	CA	ASN A			14.953	12.752	8.141		18.16
ATOM	949	C	ASN A			14.301	13.929	7.458		19.79
ATOM	950	0	ASN A			13.895	14.802	8.157		12.28
ATOM	951	CB	ASN A			16.481	12.573	8.239		14.17
ATOM	952	CG	ASN A			17.247	13.740	8.812		19.75
ATOM	953		ASN A			17.821	14.341	7.934		
ATOM	954		ASN A			17.390	14.341	10.042		14.52
ATOM	955	N	LEU A			14.180	14.130	6.141		17.43 27.31
ATOM	956	CA	LEU A			13.640	15.270	5.553		
ATOM	957	C	LEU A			12.190	15.270			25.53
ATOM	958	0	LEU A			11.710	16.281	5.971 6.549		22.45
ATOM	959	CB	LEU A			13.632	15,269			25.13
		Q <i>D</i>		100		13.032	13,209	4.056	1.00	41.28

ATOM	960	CG	LEU A	138	13.713	16.582	3.303	1.00	31.76
ATOM	961	CD1			14.641	17.503	4.012		51.09
ATOM	962	CD2	LEU A	138	14.207	16.573	1.958	1.00	
ATOM	963	N	GLN A	139	11.378	14.403	5.569	1.00	
ATOM	964	CA	GLN A		10.034	14.390	6.037	1.00	
ATOM	965	С	GLN A		9.846	14.749	7.471	1.00	
ATOM	966	0	GLN A		8.791	15.282	7.528	1.00	
ATOM	967	CB	GLN A		9.517	12.969	5.899	1.00	
ATOM	968	CG	GLN A		9.684	12.643	4.450	1.00	
ATOM	969	CD	GLN A		10.984	11.983	4.110	1.00	
ATOM	970	OE1			10.674	10.980	3.477	1.00	
ATOM	971	NE2			12.195	12.405	4.410		31.70
ATOM	972	N	ASN A		10.454	14.072	8.427	1.00	
ATOM	973	CA	ASN A		10.215	14.183	9.848		
ATOM	974	C	ASN A		10.941	15.429		1.00	
ATOM	975	Ō	ASN A		11.040	15.654	10.293	1.00	
ATOM	976	CB	ASN A		10.581	12.910	11.454 10.541	1.00	18.05
ATOM	977	CG	ASN A		9.465	11.998		1.00	
ATOM	978	OD1			8.615	12.565	10.210	1.00	
ATOM	979	ND2			9.460		9.563		23.57
ATOM	980	N	ARG A		11.457	10.756	10.630		22.65
ATOM	981	CA	ARG A		12.170	16.162	9.397		19.20
ATOM	982	C	ARG A		13.219	17.350	9.790		26.25
ATOM	983	Ö	ARG A		13.219	17.090	10.818		25.06
ATOM	984	СВ	ARG A		11.123	17.928	11.649		27.60
ATOM	985	CG	ARG A		10.083	18.299	10.271		37.72
ATOM	986	N	GLY A		14.110	18.974	9.372		49.61
ATOM	987	CA	GLY A			16.165	10.920		19.42
ATOM	988	C	GLY A		14.997	15.778	11.902		14.21
ATOM	989	Õ	GLY A		14.652	15.066	13.158		19.42
ATOM	990	N	GLY A		15.547	14.759	13.971		23.74
ATOM	991	CA	GLY A		13.354	14.851	13.569		14.09
ATOM	992	C	GLY A		13.210	14.075	14.757		11.80
ATOM	993	Ö	GLY A		12.203	12.972	14.555		16.69
ATOM	994	N	ILE A		11.760	12.787	13.481		19.57
ATOM	995	CA	ILE A		11.668	12.386	15.590		19.71
ATOM	996	C	ILE A		10.494	11.589	15.667		20.13
ATOM	997	0	ILE A		9.313	12.315	16.296		27.00
ATOM	998	CB	ILE A		9.298	13.026	17.268		26.75
ATOM	999	CG1	ILE A		10.973	10.583	16.692	1.00	
ATOM	1000	CG2	ILE A		12.363	9.956	16.348	1.00	5.60
ATOM	1000	CD1	ILE A		9.882	9.636	16.775		14.01
ATOM	1001	N	PRO A		12.437	9.156	17.562	1.00	2.75
ATOM	1002	CA	PRO A		8.249	12.380	15.499		32.77
ATOM	1003	CA	PRO A		6.959	12.993	15.779	1.00	
ATOM	1004	0	PRO A		6.484	12.588	17.180	1.00	
MOTA	1005	CB	PRO A		6.475	11.446	17.537	1.00	
ATOM	1000	CG			5.957	12.384	14.784	1.00	
111 OF1	1007	CG	PRO A	140	6.887	12.059	13.668	1.00	25.85

ATOM	1008	CD	PRO	Α	145	8	3.174	11.5	63	14.234	1.00	31.33
ATOM	1009	N	ASN	Α	146	į	5.796	13.4	62	17.878	1.00	27.07
ATOM	1010	CA			146	5	5.454	13.2	74	19.230	1.00	28.59
MOTA	1011	С			146	6	5.526	12.6	05	20.045	1.00	29.25
ATOM	1012	0	ASN	A	146	(5.087	11.9	95	20.996	1.00	35.51
ATOM	1013	CB	ASN	A	146	4	1.285	12.3	64	19.230	1.00	41.13
ATOM	1014	CG			146	3	3.300	12.5	68	18.120	1.00	48.43
MOTA	1015		ASN			3	3.134	13.7	21	17.788	1.00	49.24
ATOM	1016	ND2				2	2.763	11.4	37	17.695	1.00	47.79
MOTA	1017	N	TYR	Α	147	7	7.791	12.7	99	19.885	1.00	23.88
ATOM	1018	CA	TYR	Α	147		3.689	12.3	39	20.969	1.00	21.90
ATOM	1019	С	TYR	Α	147	ç	.583	13.4	95	21.285	1.00	22.57
MOTA	1020	0	TYR	Α	147	٥	777	14.3	99	20.494	1.00	26.53
MOTA	1021	CB	TYR	Α	147	9	3.309	11.0	98	20.498	1.00	21.16
MOTA	1022	CG			147	10	.285	10.4	71	21.349	1.00	20.45
MOTA	1023	CD1			147	9	.882	9.7	20	22.384	1.00	24.28
ATOM	1024	CD2		Α	147	11	.608	10.5	64	21.189	1.00	17.96
ATOM	1025	CE1				10	.681	9.0	29	23.273	1.00	24.55
ATOM	1026	CE2				12	.509	9.9	48	21.983	1.00	20.73
ATOM	1027	CZ	TYR			12	.022	9.1	84	23.030	1.00	24.61
MOTA	1028	OH	TYR			12	.891	8.5	36	23.887	1.00	24.80
ATOM	1029	N	PRO			9	.893	13.8	58	22.507	1.00	22.86
ATOM	1030	CA	PRO			10	.817	14.9	16	22.769	1.00	21.77
ATOM	1031	С	PRO			12	.127	14.8	82	21.957	1.00	22.49
ATOM	1032	0	PRO			13	.007	14.0	04	22.117	1.00	22.31
ATOM	1033	CB	PRO				.185	14.6	94	24.251	1.00	23.23
ATOM	1034	CG	PRO			10	.324	13.5	76	24.719	1.00	23.39
ATOM	1035	CD	PRO			9	.677	12.8		23.590	1.00	25.33
ATOM	1036	N	ARG			12	.432	15.9	80	21.250	1.00	25.45
ATOM	1037	CA	ARG			13	.735	16.1	38	20.567	1.00	22.54
ATOM	1038	С	ARG			14	.910	16.0	18	21.499	1.00	21.28
ATOM	1039	0	ARG			15	.860	15.4	77	21.015	1.00	16.61
ATOM	1040	СВ	ARG			13	.829	17.3	46	19.727	1.00	31.02
ATOM	1041	CG	ARG			12	.837	17.7	50	18.719	1.00	58.26
ATOM	1042	CD	ARG			13	.452	18.6		17.658	1.00	80.58
ATOM	1043	NE	ARG			13	.769	17.7	98	16.491	1.00	92.05
ATOM	1044	CZ	ARG			13	.315	18.1	54	15.320	1.00	91.85
ATOM	1045		ARG			12	.586	19.2		15.165	1.00	86.98
ATOM	1046		ARG			13	.544	17.48	88	14.242	1.00	91.61
ATOM	1047	N	GLU			14	.813	16.28	82	22.825	1.00	28.09
ATOM	1048	CA	GLU			15	.950	16.1	71	23.735	1.00	25.55
ATOM	1049	С	GLU				.272	14.73	36	24.020	1.00	21.12
ATOM	1050	0	GLU				.372	14.4	43	24.371	1.00	24.39
ATOM	1051	CB	GLU				.753	17.04	40	24.917	1.00	38.73
ATOM	1052	CG	GLU				.328	17.3		25.359	1.00	67.27
ATOM	1053	CD	GLU				.252	17.18	35	26.899	1.00	85.05
ATOM	1054		GLU				.005	17.89		27.657	1.00	90.70
ATOM	1055	OE2	GLU	A	150	13	.454	16.32	21	27.373	1.00	91.68

ATOM	1056	N	ND C	. 70	161	15 006			
					151	15.396	13.807	23.727	1.00 19.70
ATOM	1057	CA			151	15.752	12.424	23.844	1.00 19.52
ATOM	1058	С			151	16.163	11.779	22.531	1.00 19.28
ATOM	1059	0			151	16.373	10.586	22.480	1.00 14.55
ATOM	1060	CB			151	14.548	11.796	24.412	1.00 23.06
ATOM	1061	CG			151	13.853	12.432	25.516	1.00 22.24
ATOM	1062	CD	ARG	Α	151	13.200	11.451	26.393	1.00 33.40
ATOM	1063	NE	ARG	Α	151	12.609	11.893	27.633	1.00 46.53
ATOM	1064	CZ	ARG	Α	151	11.796	11.028	28.275	1.00 52.87
ATOM	1065	NH1			151	11.428	9.823	27.930	1.00 51.02
ATOM	1066	NH2			151	11.203	11.278	29.416	1.00 59.98
ATOM	1067	N			152	16.360	12.526	21.505	1.00 14.12
ATOM	1068	CA			152	16.629	11.925	20.253	1.00 14.12
ATOM	1069	C			152	17.995			
ATOM	1070	Ö			152		12.249	19.745	1.00 17.30
ATOM	1070	СВ				18.282	13.373	19.965	1.00 21.34
ATOM	1071	OG1			152 152	15.680	12.408	19.158	1.00 13.91
ATOM	1072	CG2				14.423	12.256	19.858	1.00 23.92
ATOM					152	15.737	11.934	17.759	1.00 6.77
	1074	N			153	18.704	11.336	19.121	1.00 15.49
ATOM	1075	CA			153	19.930	11.725	18.450	1.00 17.73
ATOM	1076	C			153	19.893	11.035	17.073	1.00 18.41
ATOM	1077	0			153	19.866	9.800	17.121	1.00 16.04
ATOM	1078	CB			153	21.112	11.260	19.338	1.00 14.55
ATOM	1079	CG	LYS			22.523	11.508	18.933	1.00 11.95
ATOM	1080	ÇD	LYS			22.883	12.882	19.403	1.00 40.35
ATOM	1081	CE	LYS			24.358	13.093	19.079	1.00 62.12
ATOM	1082	NZ	LYS			24.930	14.235	19.863	1.00 73.03
ATOM	1083	N	VAL			19.910	11.962	16.136	1.00 15.86
MOTA	1084	CA	VAL	Α	154	20.031	11.508	14.730	1.00 15.79
MOTA	1085	С	VAL	Α	154	21.406	11.481	14.040	1.00 13.11
ATOM	1086	0	VAL	Α	154	21.958	12.460	13.675	1.00 13.51
ATOM	1087	CB	VAL	Α	154	19.095	12.257	13.674	1.00 5.90
ATOM	1088	CG1	VAL	A	154	19.276	11.765	12.247	1.00 8.45
ATOM	1089	CG2	VAL	A	154	17.672	12.091	14.117	1.00 7.14
ATOM	1090	N	PHE	Α	155	22.039	10.448	13.605	1.00 13.75
ATOM	1091	CA	PHE			23.263	10.473	12.843	1.00 10.67
ATOM	1092	С			155	22.906	10.406	11.402	1.00 11.64
ATOM	1093	0	PHE			22.505	9.367	10.893	1.00 15.09
ATOM	1094	CB	PHE			23.955	9.120	13.304	1.00 15.09
ATOM	1095	CG	PHE			24.396	9.266	14.739	1.00 16.52
ATOM	1096		PHE			23.678	8.642	15.696	1.00 18.32
ATOM	1097		PHE			25.503			
ATOM	1098	CE1	PHE				9.950	15.107	1.00 11.27
ATOM	1099		PHE			24.037	8.702	17.011	1.00 23.25
ATOM	1100	CZ	PHE			25.888	9.994	16.372	1.00 7.37
ATOM	1101	N N	CYS			25.139	9.384	17.357	1.00 16.13
ATOM	1101					23.205	11.255	10.511	1.00 12.38
		CA	CYS			22.847	11.443	9.114	1.00 11.64
ATOM .	1103	С	CYS	A	126	24.057	12.027	8.461	1.00 10.08

MOTA	1104	0	CYS A	156	24.385	13.174	8.378	1.00 13.73
MOTA	1105	CB	CYS A	156	21.575	12.391	8.917	1.00 6.30
MOTA	1106	SG	CYS A	156	20.137	11.470	8.287	1.00 10.60
ATOM	1107	N	ASN A	. 157	24.814	11.147	7.918	1.00 16.95
ATOM	1108	CA	ASN A	157	26.229	11.665	7.576	1.00 19.16
ATOM	1109	С	ASN A	157	26.197	12.367	6.310	1.00 17.70
ATOM	1110	0	ASN A		25.368	12.330	5.469	1.00 20.91
ATOM	1111	CB	ASN A		27.115	10.714	8.300	1.00 30.34
ATOM	1112	CG	ASN A		27.733	9.498	7.932	1.00 34.95
ATOM	1113		ASN A		28.011	8.573	8.606	1.00 44.28
ATOM	1114		ASN A		27.965	9.541	6.660	1.00 54.18
ATOM	1115	N	VAL A		26.849	13.501	6.313	1.00 25.65
ATOM	1116	CA	VAL A		26.825	14.483	5.192	1.00 28.21
ATOM	1117	C	VAL A		26.768	13.893	3.758	1.00 24.85
ATOM	1118	Ö	VAL A		25.732	14.266	3.111	1.00 30.96
ATOM	1119	CB	VAL A		27.954	15.512	5.217	1.00 30.90
ATOM	1120	CG1			28.751	14.595	4.238	1.00 27.87
ATOM	1121		VAL A		27.791	16.704	4.236	1.00 40.31
ATOM	1122	N	GLY A		27.791	12.956		
ATOM	1123	CA	GLY A		26.713	12.774	3.016	1.00 5.94
ATOM	1124	C	GLY A		25.734		1.732	1.00 6.20
ATOM	1125	0	GLY A			11.797	1.487	1.00 4.00
ATOM	1126	N	ASP A		25.732	10.704	0.848	1.00 4.06
ATOM	1127	CA.	ASP A		25.052	11.441	2.643	1.00 8.53
ATOM	1128	CA			24.106	10.302	2.828	1.00 11.97
			ASP A		22.755	10.698	2.177	1.00 14.44
ATOM	1129	0	ASP A		21.928	11.398	2.692	1.00 10.21
ATOM	1130	CB	ASP A		24.037	9.829	4.277	1.00 12.43
ATOM	1131	CG	ASP A		23.126	8.629	4.261	1.00 20.99
ATOM	1132		ASP A		22.525	8.408	3.179	1.00 33.03
ATOM	1133		ASP A		22.956	7.840	5.216	1.00 10.13
ATOM	1134	N	ALA A		22.455	10.402	0.961	1.00 12.33
ATOM	1135	CA	ALA A		21.318	10.743	0.269	1.00 11.01
ATOM	1136	С	ALA A		19.961	10.317	0.848	1.00 15.22
ATOM	1137	0	ALA A		18.969	11.034	0.594	1.00 9.50
ATOM	1138	CB	ALA A		21.365	10.334	-1.172	1.00 13.68
MOTA	1139	N	VAL A		19.915	9.468	1.840	1.00 14.54
MOTA	1140	CA.	VAL A		18.653	9.014	2.287	1.00 9.86
MOTA	1141	С	VAL A		18.235	10.063	3.258	1.00 13.50
MOTA	1142	0	VAL A		17.094	10.458	3.377	1.00 20.47
MOTA	1143	CB	VAL A		18.596	7.778	3.117	1.00 7.34
ATOM	1144	CG1			18.931	6.592	2.259	1.00 6.50
MOTA	1145	CG2	VAL A		19.514	7.858	4.210	1.00 18.46
MOTA	1146	N	CYS A		19.198	10.733	3.719	1.00 13.44
MOTA	1147	CA	CYS A		18.864	11.811	4.720	1.00 11.26
MOTA	1148	С	CYS A		18.256	12.963	4.042	1.00 15.57
MOTA	1149	0	CYS A		18.219	13.857	4.880	1.00 14.09
MOTA	1150	CB	CYS A		20.144	12.145	5.570	1.00 18.70
MOTA	1151	SG	CYS A	163	20.748	10.705	6.581	1.00 13.38

MOTA	1152	N	THR	A	164	18.100	13.014	2.696	1.00	21.82
ATOM	1153	CA	THR	Α	164	17.603	14.283	2.171	1.00	23.08
ATOM	1154	С	THR	A	164	16.597	14.022	1.098	1.00	23.39
MOTA	1155	0	THR	A	164	16.517	14.727	0.137	1.00	33.37
MOTA	1156	CB	THR	Α	164	18.463	15.341	1.454	1.00	23.25
MOTA	1157	OG1	THR	Α	164	19.486	14.707	0.674	1.00	23.21
MOTA	1158	CG2	THR	Α	164	18.958	16.261	2.491	1.00	37.71
MOTA	1159	N	GLY	A	165	15.802	13.085	1.309	1.00	24.23
MOTA	1160	ca	GLY	A	165	14.606	12.783	0.579	1.00	26.69
MOTA	1161	С	GLY	Α	165	14.699	11.814	-0.515	1.00	28.56
ATOM	1162	0	GLY	A	165	13.680	11.775	-1.124	1.00	39.76
MOTA	1163	N	THR	A	166	15.661	11.044	-0.736	1.00	25.80
ATOM	1164	CA	THR	Α	166	16.006	10.220	-1.774	1.00	25.53
ATOM	1165	С	THR	A	166	16.195	8.866	-1.175	1.00	25.35
ATOM	1166	0	THR	Α	166	16.913	8.760	-0.206	1.00	30.91
ATOM	1167	CB	THR	Α	166	17.406	10.657	-2.230	1.00	31.57
ATOM	1168	OG1	THR	Α	166	17.105	11.788	-2.982	1.00	24.13
ATOM	1169	CG2	THR	Α	166	18.061	9.559	-2.983	1.00	34.67
ATOM	1170	N	LEU	Α	167	15.734	7.833	-1.817	1.00	19.63
ATOM	1171	CA	LEU	Α	167	16.219	6.552	-1.465	1.00	16.11
ATOM	1172	С	LEU	A	167	17.395	6.044	-2.300	1.00	
ATOM	1173	0	LEU	A	167	17.265	4.869	-2.612	1.00	
ATOM	1174	CB	LEU	Α	167	15.086	5.624	-1.555		23.45
ATOM	1175	CG	LEU	Α	167	14.123	5.773	-0.401		33.91
ATOM	1176	CD1	LEU	A	167	12.969	4.908	-0.793		42.10
ATOM	1177	CD2	LEU	A	167	14.776	5.385	0.903	1.00	
ATOM	1178	N	ILE	Α	168	18.534	6.726	-2.507		21.67
ATOM	1179	CA	ILE	Α	168	19.608	6.051	-3.170	1.00	23.38
ATOM	1180	C	ILE	Α	168	20.675	5.585	-2.189		20.47
ATOM	1181	0	ILE	Α	168	21.139	6.541	-1.581	1.00	
ATOM	1182	CB	ILE	A	168	20.254	6.835	-4.297	1.00	
ATOM	1183	CG1	ILE	Α	168	21.232	7.874	-3.800	1.00	
ATOM	1184	CG2	ILE	Α	168	19.445	7.627	-5.276	1.00	
ATOM	1185	CD1			168	20.908	8.938	-4.804	1.00	
MOTA	1186	N	ILE	A	169	21.396	4.478	-2.394	1.00	18.32
ATOM	1187	CA	ILE	A	169	22.554	4.448	-1.536		13.25
ATOM	1188	С	ILE	A	169	23.924	4.662	-1.967		11.95
ATOM	1189	0	ILE	A	169	24.615	3.942	-2.539		20.35
ATOM	1190	CB	ILE	A	169	22.503		-0.499		21.07
ATOM	1191	CG1	ILE	A	169	23.398		-0.655		11.06
ATOM	1192	CG2	ILE	Α	169	21.122	2.801	-0.533		7.02
ATOM	1193	CD1	ILE			22.581	1.266	-1.587		32.83
ATOM	1194	N	THR			24.570	5.586	-1.296		17.16
ATOM	1195	CA	THR	A	170	25.883		-1.397		13.01
ATOM	1196	С	THR			26.722		-0.240		10.14
ATOM	1197	0	THR			26.334		0.758	1.00	
ATOM	1198	CB	THR			25.623				15.02
ATOM	1199		THR			26.466	7.947	-0.255		23.39
								-	-	

ATOM	1200	CG2	THR	A	170	24.389	7.914	-0.452	1.00	41.10
ATOM	1201	N			171	28.000	5.738	-0.469	1.00	10.12
ATOM	1202	CA			171	29.012	5.066	0.339	1.00	11.88
ATOM	1203	С			171	28.897	5.492	1.765	1.00	9.74
ATOM	1204	0	PRO	Α	171	28.904	4.682	2.646	1.00	9.54
ATOM	1205	CB	PRO	Α	171	30.414	5.207	-0.286	1.00	7.15
ATOM	1206	CG	PRO	Α	171	30.017	5.603	-1.654	1.00	7.18
ATOM	1207	CD	PRO	Α	171	28.667	6.233	-1.601	1.00	6.90
ATOM	1208	N	ALA	Α	172	28.725	6.718	1.980	1.00	6.71
ATOM	1209	CA	ALA	A	172	28.247	7.315	3.169	1.00	8.62
ATOM	1210	С	ALA	Α	172	27.075	6.631	3.892	1.00	
ATOM	1211	0	ALA	Α	172	27.037	6.755	5.165		16.49
ATOM	1212	CB	ALA	Α	172	27.904	8.812	3.040	1.00	2.86
ATOM	1213	N	HIS	Α	173	26.287	5.815	3.278	1.00	6.36
ATOM	1214	CA	HIS	A	173	25.133	5.468	4.081	1.00	5.29
ATOM	1215	С	HIS	Α	173	25.685	4.314	4.888		10.58
ATOM	1216	0	HIS	Α	173	25.082	3.598	5.668	1.00	9.36
ATOM	1217	CB	HIS	Α	173	24.081	4.883	3.216	1.00	8.41
ATOM	1218	CG	HIS	Α	173	22.815	4.403	3.791	1.00	7.30
ATOM	1219	ND1	HIS	Α	173	22.066	5.327	4.565	1.00	8.48
ATOM	1220		HIS			22.148	3.264	3.670	1.00	7.83
ATOM	1221	CE1	HIS	Α	173	20.932	4.657	4.861	1.00	17.36
ATOM	1222	NE2	HIS	Α	173	20.945	3.423	4.379	1.00	5.29
ATOM	1223	N	LEU	Α	174	26.823	3.947	4.326	1.00	8.03
ATOM	1224	CA	LEU			27.344	2.623	4.682	1.00	8.06
MOTA	1225	С	LEU			28.171	2.787	5.930	1.00	13.06
ATOM	1226	0	LEU			28.609	1.648	6.151	1.00	19.88
ATOM	1227	CB	LEU			28.078	2.118	3.488	1.00	2.76
ATOM	1228	CG	LEU			27.560	0.902	2.847	1.00	13.35
ATOM	1229	CD1				26.024	1.017	2.796	1.00	18.01
MOTA	1230	CD2	LEU			27.913	0.740	1.421	1.00	21.70
ATOM	1231	N	SER			28.290	3.989	6.447	1.00	12.43
ATOM	1232	CA	SER			29.230	4.052	7.553	1.00	18.01
ATOM	1233	С	SER	A	175	28.872	4.811	8.847	1.00	19.89
MOTA	1234	0	SER			28.968	6.047	9.120	1.00	14.61
ATOM	1235	CB	SER			30.516	4.606	6.847	1.00	20.11
MOTA	1236	OG	SER			30.834	5.907	7.293	1.00	27.73
ATOM	1237	N	TYR			28.479	3.978	9.815	1.00	17.89
ATOM	1238	CA	TYR			28.092	4.530	11.133	1.00	12.54
ATOM	1239	С	TYR			28.530	3.671	12.272	1.00	11.16
MOTA	1240	0	TYR			27.949	3.770	13.257	1.00	7.63
MOTA	1241	CB	TYR			26.511	4.283	11.053	1.00	9.13
ATOM	1242	CG	TYR			25.831	5.525	10.029	1.00	5.03
ATOM	1243	CD1	TYR			25.874	6.923	10.425	1.00	2.75
ATOM	1244	CD2	TYR			25.152	5.022	8.980	1.00	2.18
ATOM	1245	CE1	TYR			25.287	7.754	9.633	1.00	4.25
ATOM	1246	CE2	TYR			24.649	5.981	8.085	1.00	6.77
ATOM	1247	CZ	TYR	A	1/6	24.658	7.329	8.399	1.00	6.22

ATOM	1248	OH	TYR A	176	24.074	8.375	7.635	1.00 5.76	5
MOTA	1249	N	THR A	177	29.430	2.685	12.167	1.00 10.72	
ATOM	1250	CA	THR A	177	29.797	1.854	13.284	1.00 13.31	
ATOM	1251	С	THR A		30.516	2.659	14.320	1.00 12.46	
ATOM	1252	0	THR A		30.311	2.436	15.475	1.00 13.12	
ATOM	1253	CB	THR A		30.658	0.683	12.798	1.00 3.49	
ATOM	1254	0G1			31.361	1.247	11.870	1.00 32.08	
ATOM	1255	CG2			29.675	-0.149	12.083	1.00 52.00	
ATOM	1256	N	ILE A		31.409	3.474	13.920		
ATOM	1257	CA	ILE A		32.203			1.00 10.48	
ATOM	1258	C	ILE A			4.246	14.783	1.00 15.25	
ATOM	1259	0	ILE A		31.180	5.045	15.632	1.00 16.95	
					31.092	4.774	16.851	1.00 22.68	
ATOM	1260	CB	ILE A		33.338	5.121	14.357	1.00 25.11	
ATOM	1261	CG1			34.701	4.496	14.056	1.00 25.05	
ATOM	1262	CG2			33.599	6.205	15.392	1.00 27.60	
ATOM	1263	CD1			34.553	3.006	14.071	1.00 55.86	
ATOM	1264	N	GLU A		30.218	5.799	15.178	1.00 16.34	
ATOM	1265	CA	GLU A		29.290	6.610	15.985	1.00 16.94	
ATOM	1266	С	GLU A		28.324	5.713	16.692	1.00 14.79	
ATOM	1267	0	GLU A	179	27.683	6.012	17.716	1.00 19.20	
ATOM	1268	CB	GLU A	179	28.555	7.637	15.169	1.00 21.16	,
ATOM	1269	CG	GLU A	179	28.790	7.283	13.691	1.00 50.37	
ATOM	1270	CD	GLU A	179	29.933	7.701	12.851	1.00 61.82	
ATOM	1271	OE1	GLU A	179	30.163	8.890	12.697	1.00 77.56	
ATOM	1272	OE2	GLU A	179	30.627	6.854	12.309	1.00 75.83	
ATOM	1273	N	ALA A	180	28.240	4.418	16.412	1.00 8.00	
ATOM	1274	CA	ALA A	180	27.353	3.520	17.042	1.00 14.34	
ATOM	1275	С	ALA A	180	28.048	2.991	18.280	1.00 19.53	
ATOM	1276	0	ALA A		27.397	3.142	19.265	1.00 21.17	
ATOM	1277	CB	ALA A		26.843	2.437	16.128	1.00 11.97	
ATOM	1278	N	ARG A		29.317	2.547	18.287	1.00 21.89	
ATOM	1279	CA	ARG A		29.992	1.982	19.398	1.00 21.89	
ATOM	1280	C	ARG A		30.296	3.106	20.367	1.00 10.48	
ATOM	1281	ō	ARG A		30.243	3.104			
ATOM	1282	CB	ARG A				21.639	1.00 28.53	
ATOM	1283	CG	ARG A		31.310	1.408	19.143	1.00 12.43	
ATOM	1284	CD	ARG A		31.954	0.432	20.052	1.00 45.44	
ATOM					32.596	-0.688	19.242	1.00 66.21	
	1285	NE	ARG A		33.333	-0.030	18.164	1.00 85.83	
ATOM	1286	CZ	ARG A		33.306	-0.321	16.895	1.00 91.35	
ATOM	1287	NH1			32.551	-1.320	16.530	1.00 96.98	
ATOM	1288		ARG A		34.023	0.400	16.095	1.00 92.83	
ATOM	1289	N	GLY A		30.387	4.262	19.847	1.00 13.94	
ATOM	1290	CA	GLY A		30.553	5.404	20.728	1.00 7.40	
ATOM	1291	С	GLY A		29.741	6.574	20.960	1.00 7.95	
ATOM	1292	0	GLY A			6.512	22.083	1.00 12.73	
MOTA	1293	N	GLU A		29.725	7.622	20.138	1.00 6.42	
ATOM	1294	CA	GLU A		28.816	8.775	20.405	1.00 10.04	
MOTA	1295	С	GLU A	183	27.421	8.369	20.645	1.00 14.41	

ATOM	1296	0	GLU	Α	183	26.8	46 8.530	21.749	1.00 15.43
ATOM	1297	CB	GLU	Α	183	29.0			1.00 21.24
ATOM	1298	CG	GLU	Α	183	28.0			1.00 62.21
ATOM	1299	CD	GLU	Α	183	28.2		19.141	1.00 81.34
ATOM	1300	OE1	GLU	Α	183	28.8			1.00 95.85
ATOM	1301	OE2			183	27.79	•		1.00 90.85
ATOM	1302	N	ALA	Α	184	26.7			1.00 15.56
ATOM	1303	CA			184	25.4			1.00 14.54
ATOM	1304	С			184	25.54			1.00 13.62
ATOM	1305	0			184	24.5			1.00 16.75
ATOM	1306	CB			184	25.03			1.00 9.58
ATOM	1307	N			185	26.42			1.00 9.42
MOTA	1308	CA			185	26.21			1.00 7.48
ATOM	1309	С			185	26.33			1.00 12.30
ATOM	1310	0			185	25.76			1.00 9.50
ATOM	1311	CB			185	27.13			1.00 4.60
ATOM	1312	N			186	27.27			1.00 15.54
ATOM	1313	CA			186	27.35			1.00 13.57
ATOM	1314	С			186	26.08			1.00 13.37
ATOM	1315	Ō			186	25.42			1.00 11.49
ATOM	1316	CB			186	28.48			1.00 30.29
ATOM	1317	CG			186	29.86			1.00 30.29
ATOM	1318	CD	ARG			30.98			1.00 37.13
ATOM	1319	NE	ARG			31.90			1.00 42.36
ATOM	1320	CZ	ARG			32.32			1.00 50.20
ATOM	1321	NH1	ARG			31.92		22.424	1.00 30.20
ATOM	1322	NH2	ARG			33.11			1.00 47.83
ATOM	1323	N			187	25.56			1.00 33.30
ATOM	1324	CA			187	24.19		24.426	1.00 13.48
ATOM	1325	С			187	23.18		25.182	1.00 15.40
ATOM	1326	O			187	22.37		25.102	1.00 13.32
ATOM	1327	CB			187	23.66		23.087	1.00 11.81
ATOM	1328	CG			187	22.28		23.032	1.00 11.61
ATOM	1329	CD1			187	21.98		23.391	1.00 14.04
ATOM	1330	CD2	PHE			21.18		22.564	1.00 8.47
ATOM	1331	CE1	PHE			20.69		23.353	1.00 10.34
ATOM	1332	CE2	PHE			19.89			1.00 17.42
ATOM	1333	CZ	PHE			19.66		22.924	1.00 17.42
ATOM	1334	N	LEU			23.03			1.00 15.17
ATOM	1335	CA	LEU			21.90			1.00 13.17
ATOM	1336	С	LEU			22.20		26.775	1.00 19.43
ATOM	1337	ō	LEU			21.28		27.461	1.00 19.07
ATOM	1338	СВ	LEU			21.70		24.552	1.00 18.72
ATOM	1339	CG	LEU			21.11			1.00 18.72
ATOM	1340		LEU			20.95			1.00 7.86
ATOM	1341		LEU			19.84			1.00 7.86
ATOM	1342	N	ARG			23.33			1.00 17.48
ATOM	1343	CA	ARG			23.79		28.547	1.00 17.48
		_			-	20.75		20.04/	T.00 TO.41

ATOM	1344	С	ARG	Α	189		23.353	7.039	29.321	1.00	16.87
ATOM	1345	0	ARG	Α	189		22.852	7.164	30.389		13.64
ATOM	1346	CB	ARG	Α	189		25.325	6.017	28.529		21.93
ATOM	1347	CG	ARG	Α	189		25.882	5.624	29.894		19.95
ATOM	1348	CD ·	ARG	Α	189		27.239	6.140	30.235		21.42
ATOM	1349	NE	ARG	Α	189		27.257	7.545	29.926		25.62
ATOM	1350	CZ	ARG	Α	189		28.491	7.983	29.699		29.22
ATOM	1351	NH1	ARG	Α	189		29.315	6.960	29.840		26.71
ATOM	1352	NH2	ARG	Α	189		28.780	9.210	29.383		33.27
ATOM	1353	N	ASP	Α	190		23.837	8.150	28.796		13.76
ATOM	1354	CA	ASP	Α	190	•	23.489	9.338	29.615		17.78
ATOM	1355	С	ASP	A	190		22.008	9.364	29.711		16.79
ATOM	1356	0	ASP	A	190		21.661	9.891	30.692		23.13
ATOM	1357	CB	ASP	Α	190		23.995	10.663	29.070		23.17
ATOM	1358	CG	ASP	Α	190		25.553	10.664	29.079		33.40
ATOM	1359	OD1	ASP	Α	190		26.250	9.836	29.761		22.68
ATOM	1360	OD2	ASP	Α	190		25.961	11.595	28.321		30.24
ATOM	1361	N	ARG	Α	191		21.156	9.128	28.781		21.61
ATOM	1362	CA	ARG	Α	191		19.707	9.265	28.849		20.99
ATOM	1363	С	ARG	Α	191		19.176	8.237	29.825		21.23
ATOM	1364	0	ARG	Α	191		18.327	8.515	30.651		20.98
ATOM	1365	CB	ARG				19.014	9.214	27.450	1.00	19.76
ATOM	1366	CG	ARG	Α	191		19.605	10.282	26.521	1.00	27.49
ATOM	1367	CD	ARG	Α	191		18.848	11.594	26.689	1.00	36.68
ATOM	1368	NE	ARG	Α	191		17.559	11.023	27.144	1.00	60.89
ATOM	1369	CZ	ARG				16.841	11.651	28.087	1.00	73.30
MOTA	1370		ARG				17.404	12.780	28.496	1.00	76.65
ATOM	1371	NH2	ARG	A	191		15.675	11.224	28.574	1.00	62.02
ATOM	1372	N	ILE	Α	192		19.734	7.037	29.885	1.00	21.02
ATOM	1373	CA	ILE				19.500	6.080	30.913	1.00	21.92
ATOM	1374	С	ILE				19.705	6.598	32.337	1.00	25.67
ATOM	1375	0	ILE				19.145	6.053	33.263	1.00	27.95
ATOM	1376	CB	ILE				20.289	4.775	30.750	1.00	24.23
ATOM	1377	CG1	ILE				19.770	4.215	29.475	1.00	26.91
ATOM	1378	CG2	ILE				19.923	3.983	31.951	1.00	15.15
ATOM	1379	CD1	ILE				20.418	2.954	29.019	1.00	21.07
ATOM	1380	N	ARG				20.535	7.574	32.629	1.00	28.72
ATOM	1381	CA	ARG				20.800	8.068	33.963		33.95
ATOM	1382	С	ARG				20.116	9.377	34.406	1.00	42.87
ATOM	1383	0	ARG				20.479	9.267	35.618		48.19
ATOM	1384	CB	ARG				22.298	8.179	34.167		34.19
ATOM	1385	CG	ARG				23.096	6.896	34.100		39.38
ATOM	1386	CD	ARG				24.590	7.213	34.133		65.92
ATOM	1387	NE	ARG				25.339	5.973	34.003		81.05
ATOM	1388	CZ	ARG				26.631	5.765	33.770		81.52
ATOM	1389		ARG				27.441	6.816	33.647	1.00	
ATOM	1390		ARG				27.120	4.536	33.652		74.00
ATOM	1391	OT	ARG	A	193		19.292	10.277	34.082	1.00	38.80
TER											

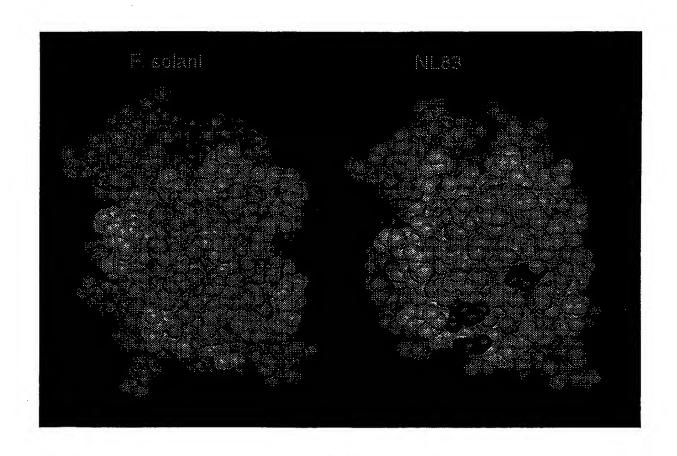


Fig. 2
3D structure of cutinases from *F. solani pisi* (left) and *H. insolens* (right)

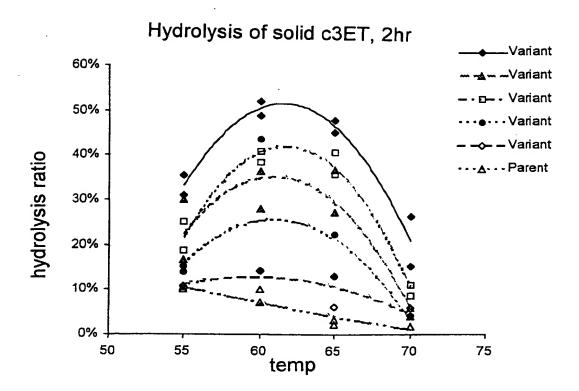


Fig. 3 Hydrolysis of solid c3ET, 2 hr

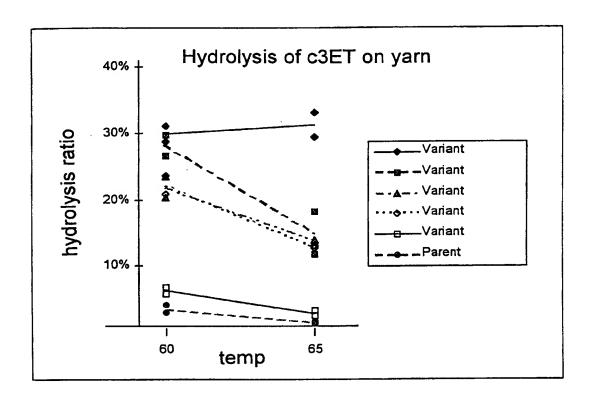


Fig. 4
Hydrolysis of c3ET on yarn, 17 hr

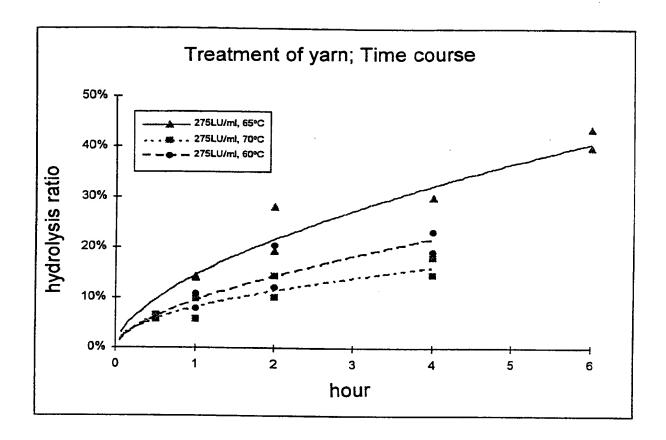


Fig. 5
Treatment of yarn; time course

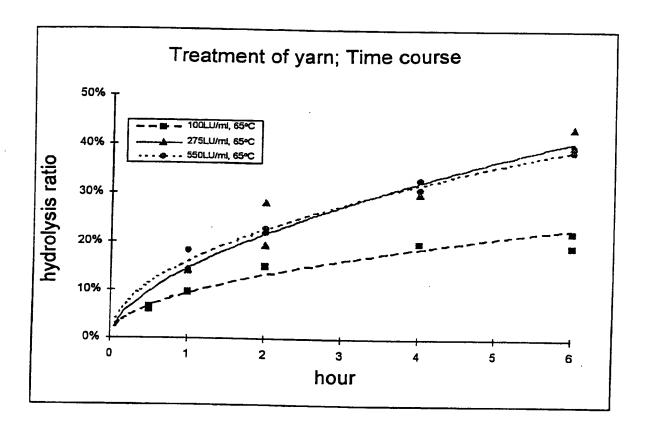


Fig. 6
Treatment of yarn; time course

International application No.

PCT/DK 99/00678

A. CLASSIFICATION OF SUBJECT MATTER

IPC7: C12N 9/18 // C11D 3/386, C08G 63/91
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC7: C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE, DK, FI, NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT
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Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 9009446 A1 (PLANT GENETICS SYSTEMS, N.V.), 23 August 1990 (23.08.90), see page 1, lines 11-20, claims	1-32,34
x	WO 9414963 A1 (UNILEVER N.V.), 7 July 1994 (07.07.94), see claim 14	1-32,34
		
A	WO 9414964 AI (UNILEVER N.V.), 7 July 1994 (07.07.94)	1-32,34
A	WO 9704078 A1 (NOVO NORDISK A/S), 6 February 1997 (06.02.97), see claim 51	1-32,34
		

X	Further documents are listed in the continuation of Box	x C. X See patent family annex.				
* "A" "E" "L" "O" "P"	Special categories of cited documents: document defining the general state of the art which is not considered to be of particular relevance erlier document but published on or after the international filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family				
	e of the actual completion of the international search May 2000	Date of mailing of the international search report 1 1 -05- 2000				
Swe	ne and mailing address of the ISA/ edish Patent Office 5055, S-102 42 STOCKHOLM simile No. +46 8 666 02 86	Authorized officer Yvonne Siösteen/EÖ Telephone No. + 46 8 782 25 00				

Form PCT/ISA/210 (second sheet) (July 1992)

International application No.
PCT/DK 99/00678

C (Continu	ation). DOCUMENTS CONSIDERED TO BE RELEVANT	PCI/DK 99/	
			In.
Category*	Citation of document, with indication, where appropriate, of the rele	vant passages	Relevant to claim No
A	PROTEINS: Structure, Function, and Genetics, Volume 26, 1996, Sonia Longhi et al, "Dyn Fusarium solani Cutinase Investigated Thr Structural Comparison Among Different Cry Forms of Its Variants" page 442 - page 45	amics of ough stal 8	1-32,34
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PCT/IS	A/210 (continuation of second sheet) (July 1992)		

International application No. PCT/DK 99/00678

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inte	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
ı. 🗀	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).:
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
	ernational Searching Authority found multiple inventions in this international application, as follows: next sheet
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-32 and 34
Remark o	on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

International application No. PCT/DK 99/00678

The invention claimed relates to two different inventions :

- Claims 1-32 and 34 relate to cutinase variants and the use of these variants.
- II. Claim 33 relates to a method for detecting cutinase activity in a sample.

Unity of invention exists only when there is a technical relationship among the claimed inventions involving one or more of the same or corresponding "special technical feature" i.e. features that define a contribution which each of the inventions make over prior art. (See Annex B to administrative instructions and Rule 13.1).